

1 *Review*

# 2 **Green Synthesis of Metal Nanoparticles from Plant** 3 **Extracts, and Their Possible Application as** 4 **Antimicrobial Agents in the Agricultural Area**

5 **Luis Castillo-Henríquez<sup>1</sup>, Karla Alfaro-Aguilar<sup>2</sup>, Jeisson Ugalde-Álvarez<sup>1</sup>, Laura**  
6 **Vega-Fernández<sup>2</sup>, Gabriela Montes de Oca-Vásquez<sup>1</sup> and José Roberto Vega-Baudrit<sup>1,2,\*</sup>**

7 <sup>1</sup> National Laboratory of Nanotechnology (LANOTEC), National Center for High Technology (CeNAT),  
8 1174-1200, San José, Costa Rica

9 <sup>2</sup> Chemistry School, National University of Costa Rica, 86-3000, Heredia, Costa Rica

10

11 \* Correspondence: jvegab@gmail.com

12 **Abstract:** Currently, metal nanoparticles have varied uses for different medical, pharmaceutical,  
13 and agricultural applications. Nano-biotechnology combined with green chemistry has great  
14 potential for the development of novel and necessary products that benefit human activities, while  
15 encourages the reduction of hazardous reagents for nanoparticle production. Green chemistry has  
16 an important role due to its contribution to unconventional synthesis methods of gold and silver  
17 nanoparticles from plant extracts, which have exhibited antimicrobial potential among other  
18 outstanding properties. Biodiversity-rich countries need to collect and convert knowledge from  
19 biological resources into processes, compounds, methods, and tools, which need to be achieved  
20 along with sustainable use and exploitation of biological diversity. Therefore, this review focuses  
21 on the importance of metal nanoparticles, the use of plant extract for their synthesis as well as other  
22 available methods, and the relevant antimicrobial activity that can be exploited in a sustainable  
23 model of agricultural management through a modern nanotechnological approach.

24 **Keywords:** agricultural industry; antibacterial; antimicrobial; green synthesis; gold;  
25 nano-biotechnology; nanoparticles; silver; sustainable development

26

## 27 **1. Introduction**

28 Currently, Green chemistry has been developed as an alternative to the use of environmentally  
29 harmful processes and products, due to the serious consequences that the world is facing, and the  
30 limited available time to find effective solutions [1-3]. According to Menges, it is suggested that  
31 green chemistry could have saved USD 65.5 billion by the end of 2020 [4].

32

33 Chen *et al.* state that circular economies should always aim to balance economic growth,  
34 resource sustainability, and environmental protection [5]. The challenge for biodiversity-rich  
35 countries and scientists is to collect and convert knowledge from biological resources into processes,  
36 compounds, methods, and tools, which need to be achieved along with sustainable use and  
37 exploitation of biological diversity [6-8]. In addition to that, biodiversity exploration has been  
38 presented to the international scientific community as a promoter of the responsible use of nature,  
39 and as a means of obtaining non-harmful components as well. For this reason, different strategies  
40 have been sought to contribute to this field through the use of green processes, such as the creation  
41 of nanoparticles (NPs) from plant extracts [9-11].

42

43 NPs are a wide range of materials with dimensions below 100 nm, which can be used in various  
44 areas such as medical, pharmaceutical, manufacturing and materials, environmental, electronics,  
45 energy collection, and mechanical industries due to their multiple properties [12-15]. In general, NPs

46 can be classified into different groups which include fullerenes, metallic NPs, ceramic NPs, and  
47 polymer NPs [15-16]. Regarding the metallic NPs, their outstanding properties have caused the  
48 development of different methodologies for their synthesis, where gold (AuNPs) and silver (AgNPs)  
49 nanoparticles prepared from plant extracts are of great interest for the researchers in their attempt to  
50 develop suitable antibacterial and antimicrobial agents for agriculture [17-20]. Also, these initiatives  
51 are considered as low-cost processes that allow avoiding toxic-generating products and benefit the  
52 agricultural activity. It is estimated that the preparation of one kg of AgNPs would cost about USD 4  
53 million, while one kg of raw silver costs around USD 14,000 [21,22].  
54

55 In 2009, Raveendran *et al.* published one of the first green synthesis methods of metal NPs. In  
56 this approach, they employed an aqueous starch solution subjected to heating, silver nitrate, and  
57 glucose as the green reducing agent [23]. After that, researchers like Iravani, and Kumar *et al.* have  
58 presented high-quality review papers regarding the synthesis of metallic nanoparticles using plant  
59 extracts as a green chemistry approach [24,25]. Since then, synthesis of metal NPs has been  
60 performed by different research groups based on a variety of plants and their structures. Logeswari  
61 *et al.* developed an eco-friendly synthesis of AgNPs from plant powders of *Solanum tuberosum*,  
62 *Syzygium cumini*, *Centella asiatica* and *Citrus sinensis*, while Yang *et al.* performed a biosynthesis of  
63 AuNPs using an agricultural waste mango peel extract [26,27]. Verma *et al.* and Bagherzade *et al.*  
64 have shown the antibacterial, and antimicrobial activity of metal NP obtained through green  
65 synthesis using *Azadirachta indica* leaves, and *Crocus sativus* L. extracts, respectively [28,29].  
66

67 Due to the nanotechnological boom, unusual physical, chemical, and biological methods have  
68 been developed for the synthesis and production of metal NPs [30-35]. Therefore, this paper seeks to  
69 describe some of these methods, the NPs' characterization techniques and also, pay particular  
70 attention to AuNPs and AgNPs' capacity as antibacterial and antimicrobial agents within the  
71 agricultural field.

## 72 2. Importance of nanoparticles

### 73 2.1. Gold Nanoparticles

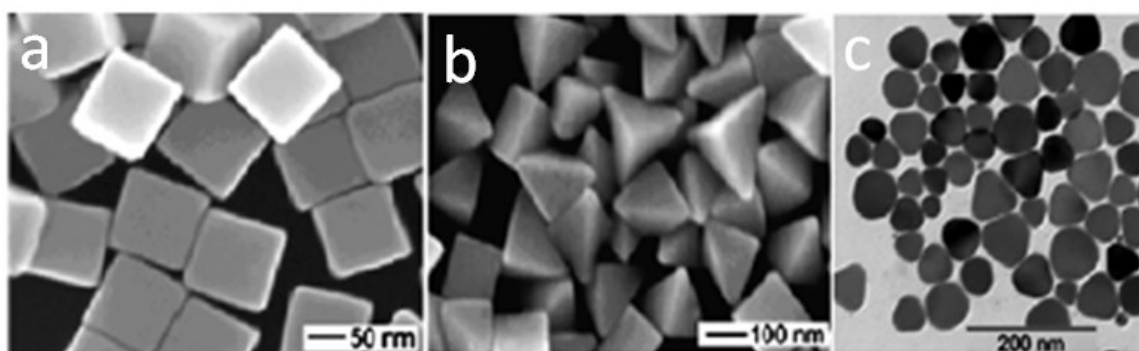
74 AuNPs can be produced in different sizes and shapes (*e.g.* nanospheres, nanocylinders,  
75 nanowires, and nanocages). AuNPs exhibit in principle, low toxicity, and multiple interesting  
76 chemical, biological, and physical properties such as photo-thermal, optical, electrochemical,  
77 biocompatibility, and they can even act as catalysts [36-38]. Also, these nanoparticles can be  
78 synthesized with ease and can fulfill relevant roles in other fields than the agricultural, like for  
79 diagnostic probes, drug development, and functionalization with a wide range of ligands such as  
80 antibodies or genetic material manipulation. Due to the previous, the demand of AuNPs is rapidly  
81 growing as a result of their outstanding properties and multiple applications [39]. The  
82 aforementioned multi-functionality represents AuNPs high scientific value and the reason why  
83 many groups are currently carrying research about them.

84 Grimaldi *et al.* present a comparison regarding two production technologies; the conventional  
85 batch production, and an innovative milli-continuous flow. The latter provides attractive features,  
86 such as high controllability of product quality, simple operation, and high efficiency in the recovery  
87 of energy, as well as in the reduction of wastes. The research reported environmental impact and  
88 costs advantages from the adoption of a continuous-flow production, instead of the conventional  
89 method. In first place, human toxicity, ecotoxicity of water, and depletion of gold resources are  
90 reduced. Additionally, the strategy implies lower costs due to milder cleaning procedures, less  
91 complex operations, reduced use of hazardous substances, and waste generation. Therefore, the  
92 proposed approach recalls the importance of recycling natural products for the production of  
93 AuNPs to avoid the depletion of natural gold resources [40].

## 94 2.2. Silver Nanoparticles

95 The synthesis AgNPs is a well-established field of work. In ancient times, silver nanoparticles  
96 were used as decorative pigments in crafts, staining glass, or ceramics. These materials have great  
97 potential and versatility as they are applicable in textiles, optoelectronics, catalysis, and  
98 environmental remediation processes as well. The latter, in particular, is due to inorganic silver's  
99 antimicrobial character, recognized as a bactericidal agent since it is an antagonist of microorganism  
100 due to its propensity for dissolution of toxic silver ion [41,42]. AgNPs show great variability in their  
101 characteristics depending on their shape and size (Figure 1). Pal *et al.* have reported that the  
102 bactericidal power of these NPs increases as they decrease in size since they have a larger surface  
103 area [43].

104 Silver has been used as a potent antimicrobial agent in different applications, having an  
105 important role in water treatment, chemical industries, food preservation, aquaculture ponds, and  
106 biomedical applications. Due to the current influence of nanotechnology, AgNPs are seen as an  
107 option towards improving agricultural productivity, through a production process that goes in  
108 harmony with the environment [44,45].



109

110 **Figure 1.** Silver nanoparticles: a) cubes, b) pyramids, and c) prisms. Adapted with permission  
111 from Marin, S. *et al.* Synthesis and characterization of silver nanoparticles and their application as an  
112 antibacterial agent. *Int J Biosen Bioelectron* 5:166-173. Copyright (2019) MedCrave [9].

## 113 3. Conventional methods for the synthesis of gold and silver nanoparticles

114 There is an incredible demand for NPs because of their fascinating properties. Due to the  
115 previous, various chemical or physical methods, and more recently biological or green  
116 chemistry-based methods have been used to streamline the process. However, among the  
117 mentioned, chemical methods are the most commonly employed, and usually have two stages:  
118 nucleation and growth. In this type of method, synthesis generally requires certain components such  
119 as a metal precursor, a reducing agent, and a stabilizing agent [46,47].

120 Physical parameters like size, size distribution, and shape can be achieved by controlling the  
121 nucleation stage, while nanoparticles growth can be controlled by adjusting experimental  
122 parameters such as the precursor used for reaction, concentration, pH, temperature, and reducing  
123 agents involved [48,49]. Cieřla *et al.* evaluated the effect of different synthesis conditions such as  
124 silver nitrate ( $\text{AgNO}_3$ ) concentration, temperature, and mechanical agitation on the properties of  
125 AgNPs. Different optical properties were observed according to NPs size and shape, as a result of  
126 the method variations tested. In this case, the mixing of reagents influenced size and shape,  
127 regardless of the process temperature. However, unmixed samples exhibited solely as spherical  
128 nanoparticles [50].

129 Chemical reduction synthesis mechanisms of AuNPs and AgNPs have been extensively used  
130 through different methods such as the Turkevich, synthesis with sodium borohydride ( $\text{NaBH}_4$ ) with

131 or without citrate, seeding-growth, synthesis by ascorbic acid, and Brust-Schiffrin [51]. Nevertheless,  
132 a major concern arises due to the use of reagents such as NaBH<sub>4</sub>, sodium citrate, ascorbate, elemental  
133 hydrogen, Tollen reagent, N, N-dimethylformamide (DMF) and block copolymers of poly (ethylene  
134 glycol) for reduction of compounds. The mentioned substances can lead to toxic by-products and  
135 damage the environment. For this reason, the use of a green pathway for the design and synthesis of  
136 NPs is currently being explored, since in this approach reducing agents are provided by plants'  
137 biomass [51,52].

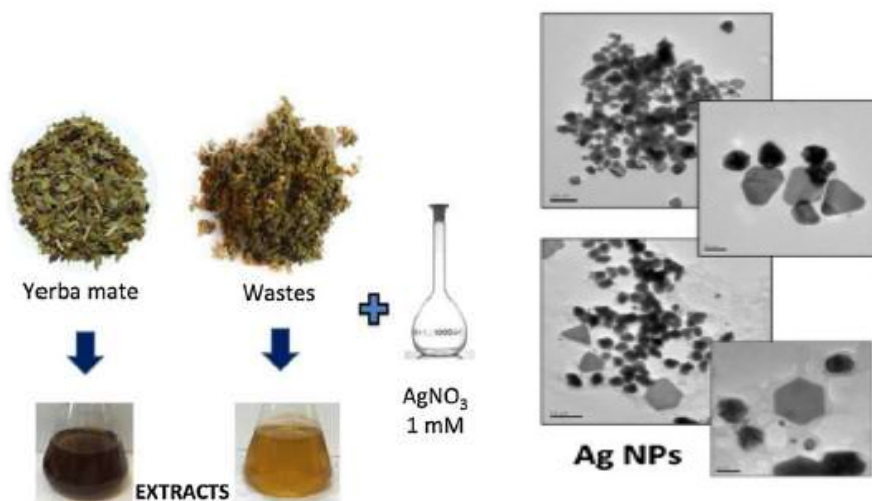
138 Although chemical methods are the most widely used and well-reported for high-quality  
139 synthesis, these may lead to NPs with a narrow size distribution, and involve the use of hazardous  
140 chemical agents (e.g. toxic organic solvents), which limit NPs applications [53]. On the other hand,  
141 physical methods include simple one-step procedures and provide large-scale production in a short  
142 time, but it is common that the resulting NPs exhibit size, shape, and size distribution defects [54].

#### 143 4. Unconventional methods for the synthesis of gold and silver nanoparticles

144 "Green methods" were created and introduced to achieve not only high social benefit but also,  
145 reducing the impact on the ecosystem. Thus, through their development and use, scientists are  
146 providing possible solutions to the issues encountered when using traditional synthesis methods,  
147 like the employment of environment-friendly solvents and reagents, as well as reducing energy  
148 consumption [55-57]. These methods consist in the use of non-toxic biomolecules such as DNA,  
149 proteins, enzymes, carbohydrates, and plant extracts for the synthesis of biocompatible metallic NPs  
150 by reducing metal ions in aqueous solutions [58,59].

151 In addition to the previous, these unconventional synthesis methods of AuNPs and AgNPs  
152 have the advantage of producing large quantities of NPs that are free from contamination and  
153 possess better-defined size and morphology than some of the obtained through conventional or  
154 physicochemical methods [60,61]. On the other hand, a disadvantage of these types of bioassays is  
155 the difficulty to establish adequate work conditions because biological raw material nature limits the  
156 set of conditions under which they can be used, and this can impact NPs formation. Therefore, it is  
157 necessary to provide well-defined specifications regarding temperature, pH, metallic solution  
158 composition, and the reaction time as well [60].

159 AgNPs synthesis through an eco-friendly and sustainable process is an important aim for  
160 nanomaterial development [62,63]. Arreche *et al.* studied two commercial brands of yerba mate (*Ilex*  
161 *paraguariensis*) for the preparation of aqueous extracts to synthesize AgNPs at room temperature  
162 (Figure 2). The obtained NPs were spherical, hexagonal, and triangular, with an average particle size  
163 of 50 nm and surface plasmon peak at 460 nm. The antimicrobial activity was evaluated against *E.*  
164 *coli* and *S. aureus*. The minimum inhibitory concentrations required for *E. coli* were 7.66 and 17.66  
165 µg\*ml<sup>-1</sup> using the treatment brand 1 and brand 2, respectively. On the other hand, the values for *S.*  
166 *aureus* were 23.25 and 50.60 µg\*ml<sup>-1</sup> for the treatment brand 1 and brand 2, respectively. The study  
167 suggested that polyphenols present in yerba mate leaf extract take action as a reducing agent and  
168 stabilizer of the nanoparticles [64].



169

170 **Figure 2.** Synthesis of AgNPs using extracts from yerba mate (*Ilex paraguariensis*) wastes.  
 171 Reprinted with permission from Arreche, R. *et al.* Synthesis of Silver Nanoparticles Using Extracts  
 172 from Yerba Mate (*Ilex paraguariensis*) Wastes. *Waste and Biomass Valorization* 73(6):1712-1720.  
 173 Copyright (2018) Springer [64].

174 Sasidharan *et al.* used the pericarp of *Myristica fragans* fruit extract for the eco-friendly synthesis  
 175 of AgNPs. In this approach, the aqueous fruit extract of the plant fulfilled reducing and stabilizing  
 176 functions for the preparation, and the obtained AgNPs exhibited good catalytic and antibacterial  
 177 activities [65]. In another approach, Alkhalaf *et al.* conducted a study to identify the effect of the  
 178 green synthesis of AgNPs from a *Nigella sativa* plant extract, resulting in NPs that exhibit antioxidant  
 179 activity [66]. Also, Sk *et al.* synthesized AuNPs and AgNPs using aqueous extract of leaves from  
 180 *Malva Verticillata*. AuNPs were found to have outstanding catalytic activity toward the hydride  
 181 transfer reduction of the aromatic nitro Schiff bases, while AgNPs displayed interesting antibacterial  
 182 activity [67].

## 183 5. Methods for obtaining plant extracts

184 Extraction methods are used for the separation of plant metabolites. In the case of AuNPs and  
 185 AgNPs synthesis, the main extraction methods employed are (a) solvent-based extraction, (b)  
 186 microwave-assisted extraction, and (c) maceration extraction [68]. The ideal extraction method  
 187 should be cost-effective, simple, less time-consuming, and carried out with ease in any laboratory  
 188 [69].

### 189 5.1. Solvent-based extraction

190 This technique allows soluble components in the solid material to be integrated with a  
 191 solvent-based extraction for mass transfer, which ratio decreases with the increase in concentration  
 192 of the soluble compound in the solvent [70]. Recently, the application of green solvents has caught  
 193 attention in different disciplines. These solvents are seen as a non-toxic, biocompatible, and  
 194 biodegradable alternatives to the conventional ones. In addition to that, they are easier to prepare  
 195 and are cost-effective. Some advances regarding green solvent technologies are deep eutectic  
 196 solvents (DESs), natural deep eutectic solvents (NDESs), ionic liquids (ILs), surfactants, and  
 197 bio-derived solvents [71,72].

### 198 5.2. Microwave-assisted extraction

199 This method employs microwave energy for the partition of analytes from the sample into the  
200 solvent by rapid heating, which allows materials to reach the necessary level of energy associated  
201 with the dielectric susceptibility of both, solvent and plant raw material [68,73]. Through its  
202 implementation; extraction time and solvent volume are reduced compared to other methods [74].  
203 The aforementioned explains why it is recognized as a green technology [75]. Aside from that,  
204 studies have shown improved recovery of analytes and reproducibility when executing the  
205 extraction by this method. However, it is necessary to take into consideration two important aspects.  
206 In first place, special concerns have to be foreseen for preventing the thermal degradation of the  
207 samples, and second, research groups need to be aware that this method is limited to small-molecule  
208 phenolic compounds [76].

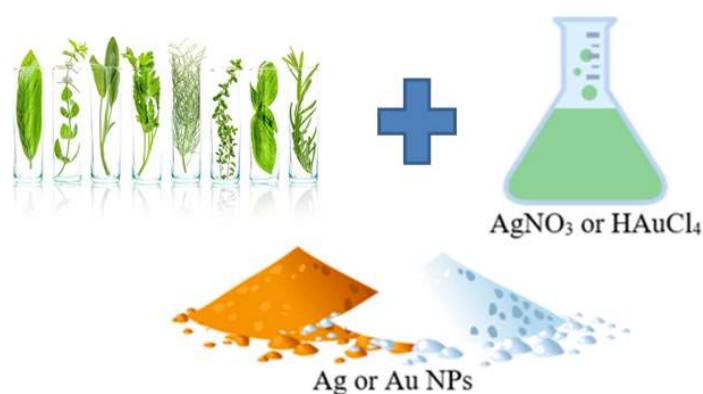
### 209 5.3. Maceration extraction

210 This process may be achieved by following three basic steps: (a) grinding the plant in small  
211 pieces, (b) adding the appropriate solvent in a closed vessel, which will determine the type of  
212 compound that is going to be extracted from the sample, and (c) filtration to separate the liquid  
213 phase [77,78]. Although this may be considered the easiest and simplest method, organic wastes can  
214 become an issue due to large amounts of solvents used, making it necessary to have a proper  
215 chemical waste management process [79].

## 216 6. Synthesis methods of gold and silver nanoparticles from plant extracts

217 The use of plant extracts is strongly arising and is conceived as a feasible alternative for the  
218 synthesis of AuNPs and AgNPs because physicochemical approaches are being considered obsolete  
219 due to costly, and hazardous materials. Plant extracts can be presented in multiple forms, and are a  
220 rich source of polyphenols, flavonoids, sugars, enzymes, and/or proteins, which can also be used as  
221 reducing and stabilizing agents for the biosynthesis of metallic NPs. A great variety of plant extracts  
222 used for generating metallic NPs have been processed and applied in various fields [80-81]. In  
223 general, this method can represent a cost-effective, environmentally-friendly, simple, and suitable  
224 option for large-scale production processes [82].

225 This method is quite diverse from others since extracts can be obtained from multiple parts of  
226 the plant or its derivative products that have demonstrated their aptitude to be considered a metal NP  
227 natural source, such as leaves, bark, stem, shoots, seeds, latex, secondary metabolites, roots, twigs,  
228 peels, fruits, seedlings, essential oils, and tissues. The extracts usually contain a large number of  
229 organic compounds in the non-volatile fraction of the active ingredients, which allows their  
230 obtention by several techniques such as extraction with polar and non-polar solvents (Figure 3) [83].



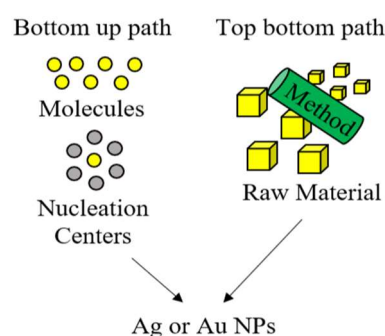
231

232

**Figure 3.** Synthesis of silver and gold nanoparticles using plant extracts.

233 Many studies have produced AgNPs from different plant extracts, such as the ones from *Citrus*  
 234 *limetta*, *Luffa acutangula*, *Parkia speciosa*, *Melia azedarach*, *Artocarpus heterophyllus*, *Azadirachta indica*,  
 235 *Gomphrena globosa*, and others [84-89]. On the other hand, different attempts have successfully  
 236 synthesized AuNPs as well, through the application of a green process such as the ones reporting the  
 237 use of *Hygrophila spinosa*, *Caulerpa racemosa*, *Eclipta alba*, *Dunaliella salina*, and *Jasminum auriculatum*  
 238 plant extracts [90-94].

239 According to Jamklände *et al.*, there are two kinds of synthesis methods depending on the  
 240 starting material for NPs preparation. In first place, the top-bottom synthesis path is employed when  
 241 raw material is at larger scales than the nano, allowing to break down its particles by grinding,  
 242 lithographic techniques, sputtering, or thermal ablation. On the other hand, the bottom-up synthesis  
 243 path includes the creation of nanoparticles from atoms that join in nucleation centers (Figure 4). In  
 244 this case, the chemical reduction processes of the compounds are fundamental [95].



245

246 **Figure 4.** Synthesis of metal NPs from Top-bottom and Bottom-up paths. Reprinted with permission  
 247 from Zhang, T. *et al.* Synthesis of Silver Nanostructures by Multistep Methods. *Sensors*  
 248 14(4):5860-5889. Copyright (2014) MDPI [52].

## 249 7. Nanoparticles characterization

250 The term characterization refers to the study of composition, structure, and other NPs properties  
 251 such as physical, chemical, electrical, and magnetic. Characterization is relevant in any study in  
 252 order to guarantee reproducible synthesis of the NPs of interest. Many techniques are currently  
 253 available for developing analytical methods for NPs characterization purposes since these possess  
 254 unique physical, chemical, and mechanical properties from bulk solids and molecules [96-98].

255 Nanomaterials have a large surface area to volume ratio, which differs greatly from the macroscopic  
 256 materials [99,100]. The physicochemical properties such as size distribution, morphology, surface  
 257 properties, chemical composition, kinetic behavior, stability, and interactions with other compounds  
 258 exhibited by AuNPs, AgNPs, and NPs in general, depend on factors like surfactant additives,  
 259 reactant concentrations, temperature, and solvent, as previously discussed. Therefore, the  
 260 nanotechnology expansion requires to use analytical techniques based on spectroscopy, diffraction,  
 261 thermal analysis, imaging, and others, for the study and characterization of nanoparticles, for which  
 262 new combinations of techniques are being developed [100-102].

### 263 7.1. Instrumental techniques for the characterization

264 Research groups have extensively used spectroscopy to detect gold and silver NPs through the  
 265 use of UV-vis methods, as these elements generate a specific signal while being reduced [103].  
 266 However, using one technique is not enough for a high-quality characterization of a sample, and  
 267 usually, a degree of uncertainty is seen in each of them [102]. Therefore, the analysis and results from  
 268 spectroscopy or any other must be supported by the combined use with other instrumental  
 269 techniques (Figure 5) [104].

270 Usually, the verification of the intended synthesis product can be done by measuring the  
271 vibrational frequencies exerted by the chemical bonds between the functional groups in the sample,  
272 which is possible thanks to Fourier transform Infrared Spectroscopy (FT-IR) when that vibrational  
273 energy is in the range of 1013-1014 Hz (*i.e.* infrared radiation). In addition to that, nanoparticles size  
274 is studied using IR, near-infrared spectroscopy (NIR), and Fourier transform near-infrared  
275 spectroscopy in the diffuse reflectance mode (DR-FTNIR) [105-107].

276 Another useful technique, X-Ray Diffraction (XRD), allows assessing the physical properties of  
277 the NPs since the diffraction pattern can give a valuable information of their average size and  
278 structure distortions of the lattice, as well as orientation. Regarding the previous, the analysis  
279 provides signals to determine whether the sample presents crystalline structures or identify the  
280 periodicity of non-crystalline amorphous phases. The two-dimensional images obtained can be  
281 converted to three-dimensional when using Fourier transform (FT) [108,109].

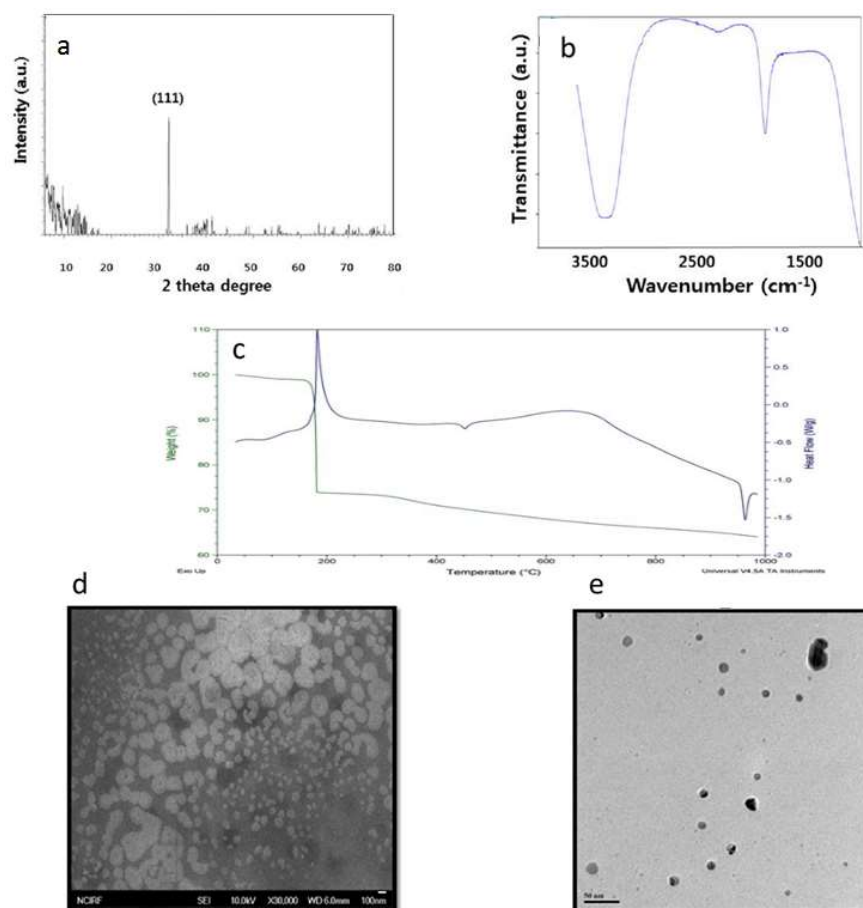
282 In addition to that, the thermal analysis performed through Differential Scanning Calorimetry  
283 (DSC), and Thermogravimetry (TG) support XRD findings. In first place, DSC thermograms enable  
284 determining if NPs possess a crystalline structure based on the form of their melting peak. Also,  
285 according to the changes from that thermal event, it is possible to determine if they interact  
286 chemically or physically with other substances. Thus, this analysis provides relevant references for  
287 chemical incompatibilities, and physical transitions such as anomerizations, crystallizations, and  
288 amorphizations [110,111]. On the other hand, TG goes into detail about mass loss of NPs due to the  
289 volatilization of one or more components such as solvents under programmed conditions of  
290 temperature to understand events like absorption, desorption, adsorption, decomposition,  
291 oxidation, and reduction [112-114].

292 Finally, Scanning electron microscopy (SEM) and Transmission electron microscopy (TEM)  
293 have been widely used for NPs characterization, where both can show the size, aggregation degree,  
294 as well as dispersion within the sample [115]. SEM is a versatile technique able to provide  
295 information about the morphology, composition, and topography of NPs surface by generating  
296 signals due to interactions presented between the electron beam and the sample [116,117].  
297 Nevertheless, TEM is considered to be the most popular technique for NPs characterization among  
298 the electron microscopy. When compared to SEM, TEM has greater capacity in providing good  
299 quality spatial resolution equal to the level of atomic dimensions, and also can perform better  
300 analytical measurements in terms of morphology, composition, and crystallographic information  
301 [117,118].

302 Studies have shown that plant extracts from different species can produce the same shaped  
303 NPs. The aforementioned is well illustrative by four studies, which reported the synthesis of  
304 spherical AgNPs through different extracts like the ones from *Tribulus Terrestris* fruit, *Alternanthera*  
305 *dentate* leaves, *Acorus calamus* roots, and *Boerhaavia diffusa* species whole plant [119-122]. On the other  
306 hand, differences in shape are expected to happen when using different structures from the same  
307 plant for the extraction process. However, as reported by Rajakumar *et al.* the use of *Eclipta prostrate*  
308 leaves for AuNPs synthesis produced NPs with triangle, pentagon, and hexagon shapes [110].

309 Green synthesis of AgNPs was performed by Katta *et al.* using *Tagetes erect* plant extract. The  
310 XRD analysis confirmed the presence of pure silver phases with a face-centered cubic structure. The  
311 UV-vis spectroscopy analysis showed an absorption peak at 420 nm, and FTIR displayed relevant  
312 vibration peaks related to silver-ion binding process and yielded polyphenols at 3401 cm<sup>-1</sup>, the  
313 presence of aromatic compounds at 2940 cm<sup>-1</sup>, and the stretching peak of C-N bond at 1104 cm<sup>-1</sup> due  
314 to the presence of plant-based amines. In addition to that, NPs morphology was assessed using SEM,  
315 where they were found to be in the range of 24-49 nm [123].





316

317 **Figure 5.** Characterization of silver nanoparticles a) XRD, b) FT-IR, c) DSC-TG, d) SEM, and e) TEM.  
 318 Reprinted with permission from Zhang, X. *et al.* Silver Nanoparticles: Synthesis, Characterization,  
 319 Properties, Applications, and Therapeutic Approaches. *Int. J. Mol. Sci* 17(9):1534-1538. Copyright  
 320 (2016) MDPI [124].

321 Furthermore, it has been found that characterization techniques may be affected by the  
 322 properties of samples regarding the material type, composition, dimensions, and the environment  
 323 where the study is conducted. For that reason, novel and sophisticated combinations of different  
 324 techniques are being developed to characterize NPs, and overcome the identified limitations [125].

### 325 8. Gold and Silver nanoparticles applications in agroindustry:

326 In general, the synthesis of NPs is of great interest because of their unique properties which can  
 327 be incorporated into composite fibers, biosensor materials, cryogenic super-conducting materials,  
 328 cosmetic products, and electronic components [126]. However, due to climate change, and the  
 329 depletion of natural resources, the synthesis of AuNPs and AgNPs from plant extracts is a major  
 330 topic for encouraging sustainable development. Because plants are the basis of this kind of green  
 331 synthesis, the created AuNPs and AgNPs can be used in many agroindustry-related processes, from  
 332 the application in the soil to the food chain [127-129].

333 Applications of nanotechnology in the food and agriculture sectors were proclaimed in June  
 334 2009 in a joint venture of the FAO and World Health Organization (WHO), with the inclusion of  
 335 wide-ranging fields, such as nanostructured ingredients, nanosized biofortification, food packaging,  
 336 nanocoating, and nanofiltration [130]. NPs may also act as “magic bullets”, containing nutrients or  
 337 other substances such as beneficial genes, and organic substances, which are targeted to specific

338 plant areas or structures to enhance their productivity. Thus, NPs represent smart nano-delivery  
339 systems for agricultural administration, specifically on crop nutrition [131].

340 Regarding direct applications of AuNPs and AgNPs, it was found that many of the researches  
341 on this field were focused on seed germination, root elongation, and plant responses towards the  
342 presence of metal NPs, like cellular oxidative stress or cytotoxicity [127,132]. In addition to that,  
343 metal NPs also have functions like nano-fertilizers, and nano-pesticides [133]. However, there are  
344 indirect applications of these NPs in the areas of food packaging, based on the antimicrobial and  
345 antibacterial activity of the AuNPs, and AgNPs [131].

#### 346 8.1. Antimicrobial and antibacterial properties.

347 In the case of metal NPs, it is well known that they can be used as antioxidants, biosensors, and  
348 for heavy metal detection [134-136]. Moreover, their unique physicochemical properties such as the  
349 ability to bind biomolecules, large surface area to volume ratio, high surface reactivity, easy to  
350 synthesize and characterize, reduced cytotoxicity, and their visible light extension behavior allow  
351 their use as antimicrobial agents [137-140]. However, this property is primarily due to their  
352 ultra-small size and shape (250 times smaller than bacteria), which enables an electrostatic  
353 interaction between the gold or silver from the NPs, and the negative charge on the cell wall or  
354 surface of microorganisms, leading them towards cellular death [141,142]. In addition to that, the  
355 high concentrations of steroids, sapogenins, carbohydrates, and flavonoids act as reducing agents of  
356 ions, and as cover agents, which provide high stability to AuNPs and AgNPs [143].

357 Many research papers in which AuNPs have shown promising antimicrobial activities have  
358 highlighted a majority spherical shape character of the NPs. Nevertheless, rod-shaped, triangular,  
359 hexagonal, and cubic NPs have also been found as part of the obtained mixture [144]. Thangamani *et al.*  
360 synthesized AuNPs using *Simarouba glauca* leaf extract. Size and shape of the NPs were sensitive  
361 to leaf broth concentration; particles tended to decrease in size with an increase in leaf broth  
362 concentration, while different morphologies were obtained such as a mixture of the prism and  
363 spherical like particles. Aside from that, they assessed the antimicrobial activity of the synthesized  
364 AuNPs by testing them against gram-positive and gram-negative organisms. The antimicrobial  
365 assay showed better results for *Staphylococcus aureus*, *Streptococcus mutans*, *Bacillus subtilis*, *Escherichia*  
366 *coli*, *Proteus vulgaris*, and *Klebsiella pneumonia* [145].

367 A study by Lediga *et al.* functionalized AgNPs with extracts of *S. birrea* and *E. autumnalis*, for  
368 which were found to exhibit remarkable antimicrobial properties against two gram-negative and  
369 two gram-positive bacteria. Both, the *S. birrea* and *E. autumnalis* AgNPs exhibited negligible or low  
370 toxicity [146]. In another approach, Montes de Oca *et al.* evaluated the impact of AgNPs usual  
371 concentrations in nature soils that are grown with Arabian Coffee in customary and organic  
372 operating systems. In this study, biomass, extracellular enzyme activities, and diversity of the soil  
373 microbial community were studied in a microcosm experiment as a function of time. After 7 days of  
374 incubation, the increase in the microbial biomass was found to be independent of AgNPs  
375 concentration [147]. In contrast, after 60 days, there was a decrease in gram-positive, and  
376 actinobacterial biomass in soils in all the evaluated AgNPs concentrations. The physicochemical  
377 properties of the soil and the enzymatic activities were not affected by AgNPs. Within the  
378 composition of the microbial community, only a few differences were observed in abundance  
379 relative to the phylum level and gender in the fungal community [147]. The results indicated that the  
380 environmental factors of AgNPs affect microbial biomass but had a low impact on microbial  
381 diversity, and may have a poor effect on soil biogeochemical cycles by extracellular enzyme  
382 activities [147,148].

## 383 9. NPs interactions with plants

### 384 9.1. Accumulation and harmful effects of NPs in plants and crops

385 Special attention should be paid to the interaction between NPs and plants (e.g. crops) when  
386 these materials are used within the agricultural field. Hashimoto *et al.* have found that accumulated  
387 AgNPs can translocate to roots and shoots of two terrestrial agro-crops; *Vigna unguiculata* and  
388 *Triticum aestivum*. Recently, it has been demonstrated that AgNPs under aerobic soil conditions are  
389 able to maintain their intact nature (88%), while a transformation to Ag<sub>2</sub>S also occurs in the same  
390 extension [149].

391 While it is not clear how metal NPs affect the environment, some studies reveal that plants  
392 overexposure to them may reveal pathways involved in the cytotoxicity. Proteomic studies on *Oryza*  
393 *sativa* (Asian rice) have increased protein precursors for oxidative stress tolerance, calcium  
394 regulation and signaling, apoptosis, and other kinds of damages [150]. This can be used for studying  
395 NPs limits in the environment. Also, high concentrations of silver can be overwhelming to the seed  
396 like for *A.thaliana*, which should not be exposed to AgNPs during its germination [151]. In contrast,  
397 there is no toxic effect on seed germination and root elongation of *Cucurbita pepo* (zucchini). This  
398 suggests that different mechanisms of action might occur across plant species concerning the effect  
399 on germination [152]. Furthermore, germination in *Lolium perenne*, *Hordeum vulgare*, and *Linum*  
400 *usitatissimum* showed to be affected at low concentrations of AgNPs but never fully inhibited [153].

401 Kaveh *et al.* studied the model agro crop *Arabidopsis thaliana* and reported the  
402 phytoaccumulation of AgNPs [154]. Another approach developed by Taylor *et al.* described *M. Sativa*  
403 *L.* (alfalfa) tendency to accumulate metal NPs of different sizes [155]. Also, gold is taken up in *A.*  
404 *thaliana* predominantly in an ionic form. It has been reported that AuNPs exposure results in the  
405 upregulation of plant genes causing downregulation of specific-metal transporters to reduce gold  
406 uptake [155].

407 Courtois *et al.* published an important study of the impact of silver species introduced into the  
408 soil via sewage sludge. As mentioned before, AgNPs are incorporated into many conventional and  
409 novel products due to their special physicochemical and antimicrobial properties. However, the  
410 discharge of these products into wastewater causes an accumulation of AgNPs and derivatives such as  
411 Ag<sub>2</sub>S in sewage sludge. The major concern is related to land application of sewage sludge for  
412 agricultural purposes since soils receive a great source of contamination for plants and crops. Soil  
413 exposure to metal NPs may lead to changes in microbial biomass, and can also affect plant growth  
414 causing physiological, biochemical, and molecular effects on them. Nonetheless, much is still  
415 unknown about the ecotoxicology of silver species, where several doubts are focused on the  
416 possibility of transfer along the trophic chain via accumulation in plants, and for that, research to  
417 evaluate the long-term impact of AgNPs on plants is ongoing [156].

### 418 9.2. NPs in soils

419 Needless to say, the growing use of AgNPs due to their recognized antimicrobial activity has  
420 led to their accumulation in soil ecosystems [157,158]. Although their environmental impact on the  
421 soil microbial community is a concern that is still under consideration, several authors have  
422 concluded that the toxic effects on microbial communities are highly dependent on the AgNPs  
423 concentration in the soil [159-163]. However, most studies have evaluated AgNPs at higher levels  
424 than actually occur in nature [164-167].

425 Meier *et al.* presented the concern that anthropogenic activities can disrupt soil ecosystems,  
426 resulting in the reduction of its microbial health. In order to evaluate the previous, they exposed  
427 freshly collected sandy loam soil to solutions ranging from 0-2000 mg/kg of AgNPs. After that, they  
428 expanded traditional soil microbial analysis with genomics-based tests through the measure of  
429 alterations in community taxonomic structure and function using 16S-rDNA profiling and

430 metatranscriptomics. The research group found that AgNPs affected bacterial taxonomic structure,  
431 as well as genes involved in heavy metal resistance, and also, their presence induced some toxicity  
432 response pathways to become highly upregulated [168].

433 Another study by Li *et al.* described the impact of AgNPs on the soil. Ag<sub>2</sub>S is more likely to be  
434 the form in which silver is retained in soils. They examined Ag<sub>2</sub>S retention from 11 natural different  
435 soils and discovered that more than 99% of the NPs were retained irrespective of the soil properties.  
436 Since the retention of Ag<sub>2</sub>S in soils is conceived as a critical factor for their toxicity and availability to  
437 sustain life (*e.g.* plants), the results obtained by this group can be a good approach for explaining the  
438 differences in phytoavailability exhibited by soils compared to what is established in the literature  
439 for liquid media [169].

## 440 10. Conclusions

441 Green chemistry is an innovative and growing resource in the search for more environmentally  
442 friendly processes. Using plant extracts for the synthesis of metal NPs is a recently growing area of  
443 interest due to its benefit in comparison to the traditional physicochemical methods. AuNPs and  
444 AgNPs generated by green synthesis have potential applications in agriculture and agroindustry,  
445 especially as antimicrobial agents of certain microorganisms for which their efficacy has been  
446 scientifically proven. Although recent studies suggest that environmental concentrations of AuNPs  
447 and AgNPs affect microbial biomass with low impact in their diversity, further research needs to be  
448 addressed in order to determine the effects they could produce to the soil, plants, and the  
449 environment in general due to long-term exposure.

450

451 **Conflicts of Interest:** The authors declare no conflict of interest.

## 452 References

- 453 1. de Marco, B.A.; Rechelo, B.S.; Tótolí, E.G.; Kogawa, A.C.; Salgado, H.R.N. Evolution of green  
454 chemistry and its multidimensional impacts: A review. *Saudi Pharmaceutical Journal* **2019**, *27*, 1–  
455 8, doi:10.1016/j.jsps.2018.07.011.
- 456 2. Hurst, G.A. Systems thinking approaches for international green chemistry education. *Current*  
457 *Opinion in Green and Sustainable Chemistry* **2020**, *21*, 93–97, doi:10.1016/j.cogsc.2020.02.004.
- 458 3. Falcone, P.M.; Hiete, M. Exploring green and sustainable chemistry in the context of  
459 sustainability transition: The role of visions and policy. *Current Opinion in Green and Sustainable*  
460 *Chemistry* **2019**, *19*, 66–75, doi:10.1016/j.cogsc.2019.08.002.
- 461 4. Menges, N. The Role of Green Solvents and Catalysts at the Future of Drug Design and of  
462 Synthesis. *Green Chemistry* **2017**, doi:10.5772/intechopen.71018.
- 463 5. Chen, T.-L.; Kim, H.; Pan, S.-Y.; Tseng, P.-C.; Lin, Y.-P.; Chiang, P.-C. Implementation of green  
464 chemistry principles in circular economy system towards sustainable development goals:  
465 Challenges and perspectives. *Science of The Total Environment* **2020**, *716*, 136998,  
466 doi:10.1016/j.scitotenv.2020.136998.
- 467 6. Ramón, A.J.L.; González, J.L.V. ESPECIES VEGETALES Valeriana pilosa, Hesperomeles  
468 ferruginea, Myrcianthes rhopaloides y Passiflora manicata FRENTE A MICRORGANISMOS  
469 PATÓGENOS Y FITOPATÓGENOS.
- 470 7. Loste, N.; Chinarro, D.; Gomez, M.; Roldán, E.; Giner, B. Assessing awareness of green  
471 chemistry as a tool for advancing sustainability. *Journal of Cleaner Production* **2020**, *256*, 120392,  
472 doi:10.1016/j.jclepro.2020.120392.

- 473 8. Zuin, V.G.; Stahl, A.M.; Zanotti, K.; Segatto, M.L. Green and Sustainable Chemistry in Latin  
474 America: which type of research is going on? And for what? *Current Opinion in Green and*  
475 *Sustainable Chemistry* **2020**, 100379, doi:10.1016/j.cogsc.2020.100379.
- 476 9. Vega-Baudrit, J.; Marin-Gamboa, S.; Rodríguez-Rojas, E.; Vega-Martínez, V. Synthesis and  
477 characterization of silver nanoparticles and their application as an antibacterial agent **2019**, 5,  
478 doi:10.15406/ijbsbe.2019.05.00172.
- 479 10. Vaid, P.; Raizada, P.; Saini, A.K.; Saini, R.V. Biogenic silver, gold and copper nanoparticles - A  
480 sustainable green chemistry approach for cancer therapy. *Sustainable Chemistry and Pharmacy*  
481 **2020**, 16, 100247, doi:10.1016/j.scp.2020.100247.
- 482 11. Mondal, P.; Anweshan, A.; Purkait, M.K. Green synthesis and environmental application of  
483 iron-based nanomaterials and nanocomposite: A review. *Chemosphere* **2020**, 259, 127509,  
484 doi:10.1016/j.chemosphere.2020.127509.
- 485 12. Khan, I.; Saeed, K.; Khan, I. Nanoparticles: Properties, applications and toxicities. *Arabian*  
486 *Journal of Chemistry* **2019**, 12, 908–931, doi:10.1016/j.arabjc.2017.05.011.
- 487 13. Saleh, T.A.; Gupta, V.K. Chapter 4 - Synthesis, Classification, and Properties of Nanomaterials.  
488 In *Nanomaterial and Polymer Membranes*; Saleh, T.A., Gupta, V.K., Eds.; Elsevier, 2016; pp. 83–  
489 133 ISBN 978-0-12-804703-3.
- 490 14. Guleria, A.; Neogy, S.; Raorane, B.S.; Adhikari, S. Room temperature ionic liquid assisted rapid  
491 synthesis of amorphous Se nanoparticles: Their prolonged stabilization and antioxidant  
492 studies. *Materials Chemistry and Physics* **2020**, 253, 123369,  
493 doi:10.1016/j.matchemphys.2020.123369.
- 494 15. Sudha, P.N.; Sangeetha, K.; Vijayalakshmi, K.; Barhoum, A. Chapter 12 - Nanomaterials  
495 history, classification, unique properties, production and market. In *Emerging Applications of*  
496 *Nanoparticles and Architecture Nanostructures*; Barhoum, A., Makhlof, A.S.H., Eds.; Micro and  
497 Nano Technologies; Elsevier, 2018; pp. 341–384 ISBN 978-0-323-51254-1.
- 498 16. Dolez, P.I. Chapter 1.1 - Nanomaterials Definitions, Classifications, and Applications. In  
499 *Nanoengineering*; Dolez, P.I., Ed.; Elsevier: Amsterdam, 2015; pp. 3–40 ISBN 978-0-444-62747-6.
- 500 17. Menazea, A.A.; Ahmed, M.K. Synthesis and antibacterial activity of graphene oxide decorated  
501 by silver and copper oxide nanoparticles. *Journal of Molecular Structure* **2020**, 1218, 128536,  
502 doi:10.1016/j.molstruc.2020.128536.
- 503 18. Panicker, S.; Ahmady, I.M.; Han, C.; Chehimi, M.; Mohamed, A.A. On demand release of ionic  
504 silver from gold-silver alloy nanoparticles: fundamental antibacterial mechanisms study.  
505 *Materials Today Chemistry* **2020**, 16, 100237, doi:10.1016/j.mtchem.2019.100237.
- 506 19. Acharya, A.; Pal, P.K. Agriculture nanotechnology: Translating research outcome to field  
507 applications by influencing environmental sustainability. *NanoImpact* **2020**, 19, 100232,  
508 doi:10.1016/j.impact.2020.100232.
- 509 20. Chandra, H.; Kumari, P.; Bontempi, E.; Yadav, S. Medicinal plants: Treasure trove for green  
510 synthesis of metallic nanoparticles and their biomedical applications. *Biocatalysis and*  
511 *Agricultural Biotechnology* **2020**, 24, 101518, doi:10.1016/j.bcab.2020.101518.
- 512 21. Ahmed, S.; Annu, Ikram, S.; Yudha S., S. Biosynthesis of gold nanoparticles: A green approach.  
513 *Journal of Photochemistry and Photobiology B: Biology* **2016**, 161, 141–153,  
514 doi:10.1016/j.jphotobiol.2016.04.034.

- 515 22. Mahawar, H.; Prasanna, R. Prospecting the interactions of nanoparticles with beneficial  
516 microorganisms for developing green technologies for agriculture. *Environmental*  
517 *Nanotechnology, Monitoring & Management* **2018**, *10*, 477–485, doi:10.1016/j.enmm.2018.09.004.
- 518 23. Raveendran, P.; Fu, J.; Wallen, S.L. Completely “Green” Synthesis and Stabilization of Metal  
519 Nanoparticles. *J. Am. Chem. Soc.* **2003**, *125*, 13940–13941, doi:10.1021/ja029267j.
- 520 24. Iravani, S. Green synthesis of metal nanoparticles using plants. *Green Chem.* **2011**, *13*, 2638–  
521 2650, doi:10.1039/C1GC15386B.
- 522 25. Mittal, A.K.; Chisti, Y.; Banerjee, U.C. Synthesis of metallic nanoparticles using plant extracts.  
523 *Biotechnology Advances* **2013**, *31*, 346–356, doi:10.1016/j.biotechadv.2013.01.003.
- 524 26. Logeswari, P.; Silambarasan, S.; Abraham, J. Ecofriendly synthesis of silver nanoparticles from  
525 commercially available plant powders and their antibacterial properties. *Scientia Iranica* **2013**,  
526 *20*, 1049–1054, doi:10.1016/j.scient.2013.05.016.
- 527 27. Yang, N.; WeiHong, L.; Hao, L. Biosynthesis of Au nanoparticles using agricultural waste  
528 mango peel extract and its in vitro cytotoxic effect on two normal cells. *Materials Letters* **2014**,  
529 *134*, 67–70, doi:10.1016/j.matlet.2014.07.025.
- 530 28. Verma, A.; Mehata, M.S. Controllable synthesis of silver nanoparticles using Neem leaves and  
531 their antimicrobial activity. *Journal of Radiation Research and Applied Sciences* **2016**, *9*, 109–115,  
532 doi:10.1016/j.jrras.2015.11.001.
- 533 29. Bagherzade, G.; Tavakoli, M.M.; Namaei, M.H. Green synthesis of silver nanoparticles using  
534 aqueous extract of saffron (*Crocus sativus* L.) wastages and its antibacterial activity against six  
535 bacteria. *Asian Pacific Journal of Tropical Biomedicine* **2017**, *7*, 227–233,  
536 doi:10.1016/j.apjtb.2016.12.014.
- 537 30. Sherin, L.; Sohail, A.; Amjad, U.-S.; Mustafa, M.; Jabeen, R.; Ul-Hamid, A. Facile green  
538 synthesis of silver nanoparticles using *Terminalia bellerica* kernel extract for catalytic  
539 reduction of anthropogenic water pollutants. *Colloid and Interface Science Communications* **2020**,  
540 *37*, 100276, doi:10.1016/j.colcom.2020.100276.
- 541 31. Li, M.; Yu, H.; Cheng, Y.; Guo, Y.; Yao, W.; Xie, Y. Simultaneous and rapid determination of  
542 polycyclic aromatic hydrocarbons by facile and green synthesis of silver nanoparticles as  
543 effective SERS substrate. *Ecotoxicology and Environmental Safety* **2020**, *200*, 110780,  
544 doi:10.1016/j.ecoenv.2020.110780.
- 545 32. Trotsiuk, L.; Antanovich, A.; Lizunova, A.; Kulakovich, O. Direct synthesis of amphiphilic  
546 polyvinylpyrrolidone-capped gold nanoparticles in chloroform. *Colloid and Interface Science*  
547 *Communications* **2020**, *37*, 100289, doi:10.1016/j.colcom.2020.100289.
- 548 33. Adamo, C.B.; Junger, A.S.; Bressan, L.P.; da Silva, J.A.F.; Poppi, R.J.; de Jesus, D.P. Fast and  
549 straightforward in-situ synthesis of gold nanoparticles on a thread-based microfluidic device  
550 for application in surface-enhanced Raman scattering detection. *Microchemical Journal* **2020**,  
551 *156*, 104985, doi:10.1016/j.microc.2020.104985.
- 552 34. Bandeira, M.; Giovanella, M.; Roesch-Ely, M.; Devine, D.M.; da Silva Crespo, J. Green synthesis  
553 of zinc oxide nanoparticles: A review of the synthesis methodology and mechanism of  
554 formation. *Sustainable Chemistry and Pharmacy* **2020**, *15*, 100223, doi:10.1016/j.scp.2020.100223.
- 555 35. Ohara, Y.; Akazawa, K.; Shibata, K.; Hirota, T.; Kodama, Y.; Amemiya, T.; Wang, J.;  
556 Yamaguchi, T. Seed-mediated gold nanoparticle synthesis via photochemical reaction of

- 557 benzoquinone. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* **2020**, *586*, 124209,  
558 doi:10.1016/j.colsurfa.2019.124209.
- 559 36. Al-Qadi, S.; Remuñán, C. Nanopartículas metálicas: oro. In: *Nanotecnología Farmacéutica*  
560 *realidades y posibilidades farmacoterapéuticas*; Vila, J. Real Academia Nacional de Farmacia,  
561 2009. ISBN 978-84-936890-8-7.
- 562 37. Gómez-López, P.; Puente-Santiago, A.; Castro-Beltrán, A.; Santos do Nascimento, L.A.; Balu,  
563 A.M.; Luque, R.; Alvarado-Beltrán, C.G. Nanomaterials and catalysis for green chemistry.  
564 *Current Opinion in Green and Sustainable Chemistry* **2020**, *24*, 48–55,  
565 doi:10.1016/j.cogsc.2020.03.001.
- 566 38. Kalimuthu, K.; Cha, B.S.; Kim, S.; Park, K.S. Eco-friendly synthesis and biomedical applications  
567 of gold nanoparticles: A review. *Microchemical Journal* **2020**, *152*, 104296,  
568 doi:10.1016/j.microc.2019.104296.
- 569 39. Solano-Umaña, V.; Vega-Baudrit, J. Controlled Deposition of Gold and Silver on a Porous  
570 Silicone Matrix. *Jacobs Journal of Nanomedicine and Nanotechnology* **2016**, *1*, 1-9.
- 571 40. Grimaldi, F.; Pucciarelli, M.; Gavriilidis, A.; Dobson, P.; Lettieri, P. Anticipatory life cycle  
572 assessment of gold nanoparticles production: Comparison of milli-continuous flow and batch  
573 synthesis. *Journal of Cleaner Production* **2020**, *269*, 122335, doi:10.1016/j.jclepro.2020.122335.
- 574 41. Calderón-Jiménez, B.; Johnson, M.E.; Montoro-Bustos, A.R.; Murphy, K.E.; Winchester, M.R.;  
575 Vega-Baudrit, J.R. Silver Nanoparticles: Technological Advances, Societal Impacts, and  
576 Metrological Challenges. *Front Chem* **2017**, *5*, 6, doi:10.3389/fchem.2017.00006.
- 577 42. Solano-Umaña, V.; Vega-Baudrit, J. Gold, silver, copper and silicone hybrid nanostructure  
578 cytotoxicity. *International Journal of Recent Scientific Research* **2017**, *8*, 2, 15478-15486.
- 579 43. Pal, S.; Tak, Y.; Song, J. Does the Antibacterial Activity of Silver Nanoparticles Depend on the  
580 Shape of the Nanoparticle? A Study of the Gram-Negative Bacterium *Escherichia coli*. *Applied*  
581 *and Environmental Microbiology* **2007**, *73*, 6, 1712-1720, doi:10.1128/AEM.02218-06.
- 582 44. Anjali Das, C.G.; Ganesh Kumar, V.; Stalin Dhas, T.; Karthick, V.; Govindaraju, K.; Mary  
583 Joselin, J.; Baalamurugan, J. Antibacterial activity of silver nanoparticles (biosynthesis): A short  
584 review on recent advances. *Biocatalysis and Agricultural Biotechnology* **2020**, 101593,  
585 doi:10.1016/j.bcab.2020.101593.
- 586 45. Kumar Panda, M.; Kumar Dhal, N.; Kumar, M.; Manjari Mishra, P.; Kumar Behera, R. Green  
587 synthesis of silver nanoparticles and its potential effect on phytopathogens. *Materials Today:*  
588 *Proceedings* **2020**, doi:10.1016/j.matpr.2020.05.188.
- 589 46. Hira, S.A.; Nallal, M.; Park, K.H. Fabrication of PdAg nanoparticle infused metal-organic  
590 framework for electrochemical and solution-chemical reduction and detection of toxic  
591 4-nitrophenol. *Sensors and Actuators B: Chemical* **2019**, *298*, 126861,  
592 doi:10.1016/j.snb.2019.126861.
- 593 47. Yang, X.; Xu, Q. Gold-containing metal nanoparticles for catalytic hydrogen generation from  
594 liquid chemical hydrides. *Chinese Journal of Catalysis* **2016**, *37*, 1594–1599,  
595 doi:10.1016/S1872-2067(16)62547-0.
- 596 48. Singh, R.; Nawale, L.U.; Arkile, M.; Shedbalkar, U.U.; Wadhvani, S.A.; Sarkar, D.; Chopade,  
597 B.A. Chemical and biological metal nanoparticles as antimycobacterial agents: A comparative  
598 study. *International Journal of Antimicrobial Agents* **2015**, *46*, 183–188,  
599 doi:10.1016/j.ijantimicag.2015.03.014.

- 600 49. Sadhasivam, S.; Vinayagam, V.; Balasubramaniyan, M. Recent advancement in biogenic  
601 synthesis of iron nanoparticles. *Journal of Molecular Structure* **2020**, *1217*, 128372,  
602 doi:10.1016/j.molstruc.2020.128372.
- 603 50. Cieśla, J.; Chylińska, M.; Zdunek, A.; Szymańska-Chargot, M. Effect of different conditions of  
604 synthesis on properties of silver nanoparticles stabilized by nanocellulose from carrot pomace.  
605 *Carbohydrate Polymers* **2020**, *245*, 116513, doi:10.1016/j.carbpol.2020.116513.
- 606 51. Daruich De Souza, C.; Ribeiro Nogueira, B.; Rostelato, M.E.C.M. Review of the methodologies  
607 used in the synthesis gold nanoparticles by chemical reduction. *Journal of Alloys and Compounds*  
608 **2019**, *798*, 714–740, doi:10.1016/j.jallcom.2019.05.153.
- 609 52. Zhang, T.; Song, Y.-J.; Zhang, X.-Y.; Wu, J.-Y. Synthesis of Silver Nanostructures by Multistep  
610 Methods. *Sensors (Basel)* **2014**, *14*, 5860–5889, doi:10.3390/s140405860.
- 611 53. Arya, A.; Mishra, V.; Chundawat, T.S. Green synthesis of silver nanoparticles from green algae  
612 (*Botryococcus braunii*) and its catalytic behavior for the synthesis of benzimidazoles. *Chemical*  
613 *Data Collections* **2019**, *20*, 100190, doi:10.1016/j.cdc.2019.100190.
- 614 54. Ocsoy, I.; Tasdemir, D.; Mazicioglu, S.; Tan, W. Nanotechnology in Plants. *Adv. Biochem. Eng.*  
615 *Biotechnol.* **2018**, *164*, 263–275, doi:10.1007/10\_2017\_53.
- 616 55. Nogueira, L.F.B.; Guidelli, É.J.; Jafari, S.M.; Ramos, A.P. 7 - Green synthesis of metal  
617 nanoparticles by plant extracts and biopolymers. In *Handbook of Food Nanotechnology*; Jafari,  
618 S.M., Ed.; Academic Press, 2020; pp. 257–278 ISBN 978-0-12-815866-1.
- 619 56. Hekmati, M.; Hasanirad, S.; Khaledi, A.; Esmaeili, D. Green synthesis of silver nanoparticles  
620 using extracts of *Allium rotundum* L, *Falcaria vulgaris* Bernh, and *Ferulago angulate* Boiss, and  
621 their antimicrobial effects in vitro. *Gene Reports* **2020**, *19*, 100589,  
622 doi:10.1016/j.genrep.2020.100589.
- 623 57. Khalaj, M.; Kamali, M.; Costa, M.E.V.; Capela, I. Green synthesis of nanomaterials - A  
624 scientometric assessment. *Journal of Cleaner Production* **2020**, *267*, 122036,  
625 doi:10.1016/j.jclepro.2020.122036.
- 626 58. Hou, D.; O'Connor, D. Chapter 1 - Green and sustainable remediation: concepts, principles,  
627 and pertaining research. In *Sustainable Remediation of Contaminated Soil and Groundwater*; Hou,  
628 D., Ed.; Butterworth-Heinemann, 2020; pp. 1–17 ISBN 978-0-12-817982-6.
- 629 59. Fasciotti, M. Perspectives for the use of biotechnology in green chemistry applied to  
630 biopolymers, fuels and organic synthesis: from concepts to a critical point of view. *Sustainable*  
631 *Chemistry and Pharmacy* **2017**, *6*, 82–89, doi:10.1016/j.scp.2017.09.002.
- 632 60. Arias Ortiz, J.D.; Palma Holguín, M.I. Elaboración de un compuesto antimicrobial con  
633 nanopartículas de plata sintetizadas a partir del extracto de hojas de romero (*rosmarinus*  
634 *officinalis*), para ser aplicado en frutas Frescas. Thesis, Universidad de Guayaquil, Facultad de  
635 Ingeniería Química., 2019.
- 636 61. Noah, N. Chapter 6 - Green synthesis: Characterization and application of silver and gold  
637 nanoparticles. In *Green Synthesis, Characterization and Applications of Nanoparticles*; Shukla, A.K.,  
638 Iravani, S., Eds.; Micro and Nano Technologies; Elsevier, 2019; pp. 111–135 ISBN  
639 978-0-08-102579-6.
- 640 62. Muthukumar, H.; Palanirajan, S.K.; Shanmugam, M.K.; Gummadi, S.N. Plant extract mediated  
641 synthesis enhanced the functional properties of silver ferrite nanoparticles over chemical  
642 mediated synthesis. *Biotechnology Reports* **2020**, *26*, e00469, doi:10.1016/j.btre.2020.e00469.



- 643 63. Pereira, T.M.; Polez, V.L.P.; Sousa, M.H.; Silva, L.P. Modulating physical, chemical, and  
644 biological properties of silver nanoparticles obtained by green synthesis using different parts  
645 of the tree *Handroanthus heptaphyllus* (Vell.) Mattos. *Colloid and Interface Science*  
646 *Communications* **2020**, *34*, 100224, doi:10.1016/j.colcom.2019.100224.
- 647 64. Arreche, R.A.; Montes de Oca-Vásquez, G.; Vega-Baudrit, J.R.; Vázquez, P.G. Synthesis of  
648 Silver Nanoparticles Using Extracts from Yerba Mate (*Ilex paraguariensis*) Wastes. *Waste*  
649 *Biomass Valor* **2020**, *11*, 245–253, doi:10.1007/s12649-018-0394-7.
- 650 65. Sasidharan, D.; Namitha, T.R.; Johnson, S.P.; Jose, V.; Mathew, P. Synthesis of silver and  
651 copper oxide nanoparticles using *Myristica fragrans* fruit extract: Antimicrobial and catalytic  
652 applications. *Sustainable Chemistry and Pharmacy* **2020**, *16*, 100255, doi:10.1016/j.scp.2020.100255.
- 653 66. Alkhalaf, M.I.; Hussein, R.H.; Hamza, A. Green synthesis of silver nanoparticles by *Nigella*  
654 *sativa* extract alleviates diabetic neuropathy through anti-inflammatory and antioxidant  
655 effects. *Saudi Journal of Biological Sciences* **2020**, doi:10.1016/j.sjbs.2020.05.005.
- 656 67. Sk, I.; Khan, M.A.; Haque, A.; Ghosh, S.; Roy, D.; Homechudhuri, S.; Alam, A. Synthesis of  
657 Gold and Silver Nanoparticles Using *Malva Verticillata* Leaves Extract: Study of Gold  
658 Nanoparticles Catalysed Reduction of Nitro-Schiff Bases and Antibacterial Activities of Silver  
659 Nanoparticles. *Current Research in Green and Sustainable Chemistry* **2020**,  
660 doi:10.1016/j.crgsc.2020.05.003.
- 661 68. Azwanida, N. A Review on the Extraction Methods Use in Medicinal Plants, Principle,  
662 Strength and Limitation. *Med Aromat Plants* **2015**, *4*, 1000196, doi:10.4172/2167-0412.1000196.
- 663 69. Yahya, N.A.; Attan, N.; Wahab, R.A. An overview of cosmeceutically relevant plant extracts  
664 and strategies for extraction of plant-based bioactive compounds. *Food and Bioprocess*  
665 *Processing* **2018**, *112*, 69–85, doi:10.1016/j.fbp.2018.09.002.
- 666 70. Schlosser, Š.; Kertész, R.; Marták, J. Recovery and separation of organic acids by  
667 membrane-based solvent extraction and pertraction: An overview with a case study on  
668 recovery of MPCA. *Separation and Purification Technology* **2005**, *41*, 237–266,  
669 doi:10.1016/j.seppur.2004.07.019.
- 670 71. Hashemi, B.; Zohrabi, P.; Dehdashtian, S. Application of green solvents as sorbent modifiers in  
671 sorptive-based extraction techniques for extraction of environmental pollutants. *TrAC Trends*  
672 *in Analytical Chemistry* **2018**, *109*, 50–61, doi:10.1016/j.trac.2018.09.026.
- 673 72. Sánchez-Camargo, A. del P.; Bueno, M.; Parada-Alfonso, F.; Cifuentes, A.; Ibáñez, E. Hansen  
674 solubility parameters for selection of green extraction solvents. *TrAC Trends in Analytical*  
675 *Chemistry* **2019**, *118*, 227–237, doi:10.1016/j.trac.2019.05.046.
- 676 73. Bhat, A.R.; Najjar, M.H.; Dongre, R.S.; Akhter, M.S. Microwave assisted synthesis of  
677 Knoevenagel Derivatives using water as green solvent. *Current Research in Green and Sustainable*  
678 *Chemistry* **2020**, doi:10.1016/j.crgsc.2020.06.001.
- 679 74. Lux, C.; Lubio, A.; Ruediger, A.; Robert, S.; Muehlethaler, C. Optimizing the analysis of dyes  
680 by Surface-Enhanced Raman Spectroscopy (SERS) using a conventional-microwave silver  
681 nanoparticles synthesis. *Forensic Chemistry* **2019**, *16*, 100186, doi:10.1016/j.forc.2019.100186.
- 682 75. Ahmed, S.; Ahmad, M.; Swami, B.L.; Ikram, S. A review on plants extract mediated synthesis  
683 of silver nanoparticles for antimicrobial applications: A green expertise. *Journal of Advanced*  
684 *Research* **2016**, *7*, 17–28, doi:10.1016/j.jare.2015.02.007.

- 685 76. Rao, S.S.; Saptami, K.; Venkatesan, J.; Rekha, P.D. Microwave-assisted rapid synthesis of silver  
686 nanoparticles using fucoidan: Characterization with assessment of biocompatibility and  
687 antimicrobial activity. *International Journal of Biological Macromolecules* **2020**,  
688 doi:10.1016/j.ijbiomac.2020.06.230.
- 689 77. Trusheva, B.; Trunkova, D.; Bankova, V. Different extraction methods of biologically active  
690 components from propolis: a preliminary study. *Chemistry Central Journal* **2007**, *1*, 13,  
691 doi:10.1186/1752-153X-1-13.
- 692 78. Oroian, M.; Dranca, F.; Ursachi, F. Comparative evaluation of maceration, microwave and  
693 ultrasonic-assisted extraction of phenolic compounds from propolis. *J Food Sci Technol* **2020**, *57*,  
694 70–78, doi:10.1007/s13197-019-04031-x.
- 695 79. Ferioli, F.; Giambanelli, E.; D'Alessandro, V.; D'Antuono, L.F. Comparison of two extraction  
696 methods (high pressure extraction vs. maceration) for the total and relative amount of  
697 hydrophilic and lipophilic organosulfur compounds in garlic cloves and stems. An application  
698 to the Italian ecotype "Aglio Rosso di Sulmona" (Sulmona Red Garlic). *Food Chemistry* **2020**,  
699 *312*, 126086, doi:10.1016/j.foodchem.2019.126086.
- 700 80. Ali, J.; Ali, N.; Wang, L.; Waseem, H.; Pan, G. Revisiting the mechanistic pathways for bacterial  
701 mediated synthesis of noble metal nanoparticles. *Journal of Microbiological Methods* **2019**, *159*,  
702 18–25, doi:10.1016/j.mimet.2019.02.010.
- 703 81. Nishanthi, R.; Malathi, S.; Paul, J.; Palani, P. Green synthesis and characterization of  
704 bioinspired silver, gold and platinum nanoparticles and evaluation of their synergistic  
705 antibacterial activity after combining with different classes of antibiotics. *Materials Science and*  
706 *Engineering: C* **2019**, *96*, 693–707, doi:10.1016/j.msec.2018.11.050.
- 707 82. Bhattarai, B.; Zaker, Y.; Bigioni, T.P. Green synthesis of gold and silver nanoparticles:  
708 Challenges and opportunities. *Current Opinion in Green and Sustainable Chemistry* **2018**, *12*, 91–  
709 100, doi:10.1016/j.cogsc.2018.06.007.
- 710 83. Sani, M.; Tatiana, A. Síntesis y caracterización de nanopartículas de plata a partir de varios  
711 extractos pigmentados de dos plantas para su aplicación en celdas solares híbridas. **2017**.
- 712 84. Dutta, T.; Ghosh, N.N.; Das, M.; Adhikary, R.; Mandal, V.; Chattopadhyay, A.P. Green  
713 synthesis of antibacterial and antifungal silver nanoparticles using Citrus limetta peel extract:  
714 Experimental and theoretical studies. *Journal of Environmental Chemical Engineering* **2020**, *8*,  
715 104019, doi:10.1016/j.jece.2020.104019.
- 716 85. Taruna; Kaushal, J.; Bhatti, J.; Kumar, P. Green synthesis and physico-chemical study of silver  
717 nanoparticles extracted from a natural source *Luffa acutangula*. *Journal of Molecular Liquids*  
718 **2016**, *224*, 991–998, doi:10.1016/j.molliq.2016.10.065.
- 719 86. Ravichandran, V.; Vasanthi, S.; Shalini, S.; Shah, S.A.A.; Tripathy, M.; Paliwal, N. Green  
720 synthesis, characterization, antibacterial, antioxidant and photocatalytic activity of *Parkia*  
721 *speciosa* leaves extract mediated silver nanoparticles. *Results in Physics* **2019**, *15*, 102565,  
722 doi:10.1016/j.rinp.2019.102565.
- 723 87. Jebril, S.; Khanfir Ben Jenana, R.; Dridi, C. Green synthesis of silver nanoparticles using *Melia*  
724 *azedarach* leaf extract and their antifungal activities: In vitro and in vivo. *Materials Chemistry*  
725 *and Physics* **2020**, *248*, 122898, doi:10.1016/j.matchemphys.2020.122898.

- 726 88. Manik, U.P.; Nande, A.; Raut, S.; Dhoble, S.J. Green synthesis of silver nanoparticles using  
727 plant leaf extraction of *Artocarpus heterophyllus* and *Azadirachta indica*. *Results in Materials*  
728 **2020**, *6*, 100086, doi:10.1016/j.rinma.2020.100086.
- 729 89. Tamilarasi, P.; Meena, P. Green synthesis of silver nanoparticles (Ag NPs) using *Gomphrena*  
730 *globosa* (Globe amaranth) leaf extract and their characterization. *Materials Today: Proceedings*  
731 **2020**, doi:10.1016/j.matpr.2020.04.025.
- 732 90. Satpathy, S.; Patra, A.; Ahirwar, B.; Hussain, M.D. Process optimization for green synthesis of  
733 gold nanoparticles mediated by extract of *Hygrophila spinosa* T. Anders and their biological  
734 applications. *Physica E: Low-dimensional Systems and Nanostructures* **2020**, *121*, 113830,  
735 doi:10.1016/j.physe.2019.113830.
- 736 91. Manikandakrishnan, M.; Palanisamy, S.; Vinosha, M.; Kalanjaraja, B.; Mohandoss, S.;  
737 Manikandan, R.; Tabarsa, M.; You, S.; Prabhu, N.M. Facile green route synthesis of gold  
738 nanoparticles using *Caulerpa racemosa* for biomedical applications. *Journal of Drug Delivery*  
739 *Science and Technology* **2019**, *54*, 101345, doi:10.1016/j.jddst.2019.101345.
- 740 92. Vijayakumar, S.; Vinayagam, R.; Anand, M.A.V.; Venkatachalam, K.; Saravanakumar, K.;  
741 Wang, M.-H.; Casimeer C, S.; Km, G.; David, E. Green synthesis of gold nanoparticle using  
742 *Eclipta alba* and its antidiabetic activities through regulation of Bcl-2 expression in pancreatic  
743 cell line. *Journal of Drug Delivery Science and Technology* **2020**, *58*, 101786,  
744 doi:10.1016/j.jddst.2020.101786.
- 745 93. Singh, A.K.; Tiwari, R.; Singh, V.K.; Singh, P.; Khadim, S.R.; Singh, U.; Laxmi; Srivastava, V.;  
746 Hasan, S.H.; Asthana, R.K. Green synthesis of gold nanoparticles from *Dunaliella salina*, its  
747 characterization and in vitro anticancer activity on breast cancer cell line. *Journal of Drug*  
748 *Delivery Science and Technology* **2019**, *51*, 164–176, doi:10.1016/j.jddst.2019.02.023.
- 749 94. Balasubramanian, S.; Kala, S.M.J.; Pushparaj, T.L. Biogenic synthesis of gold nanoparticles  
750 using *Jasminum auriculatum* leaf extract and their catalytic, antimicrobial and anticancer  
751 activities. *Journal of Drug Delivery Science and Technology* **2020**, *57*, 101620,  
752 doi:10.1016/j.jddst.2020.101620.
- 753 95. Jamkhande, P.G.; Ghule, N.W.; Bamer, A.H.; Kalaskar, M.G. Metal nanoparticles synthesis: An  
754 overview on methods of preparation, advantages and disadvantages, and applications. *Journal*  
755 *of Drug Delivery Science and Technology* **2019**, *53*, 101174, doi:10.1016/j.jddst.2019.101174.
- 756 96. Lin, P.-C.; Lin, S.; Wang, P.C.; Sridhar, R. Techniques for physicochemical characterization of  
757 nanomaterials. *Biotechnology Advances* **2014**, *32*, 711–726, doi:10.1016/j.biotechadv.2013.11.006.
- 758 97. Hans, J.; Bruhne, K. Carbon-based Nanomaterials and Hybrids: Synthesis, Properties, and  
759 Commercial Applications. Jenny Stanford Publishing, 2014; pp. 120-132 ISBN 9789814316859.
- 760 98. Kundu, M.; Pramanik, P.; Maity, A.; Joshi, D.C.; Wani, S.H.; Krishnan, P.; Mukherjee, A.;  
761 Shubha, K. Chapter 2 - Engineered Nanomaterials: Classification and Strategies for  
762 Physicochemical Characterization and Advanced Analytical Techniques for the Measurement  
763 of Nanomaterials in Plant Samples. In *Advances in Phytanotechnology*; Ghorbanpour, M.,  
764 Wani, S.H., Eds.; Academic Press, 2019; pp. 17–43 ISBN 978-0-12-815322-2.
- 765 99. Castillo-Henríquez, L.; Vargas-Zúñiga, R.; Pacheco-Molina, J.; Vega-Baudrit, J. Electrospun  
766 nanofibers: A nanotechnological approach for drug delivery and dissolution optimization in  
767 poorly water-soluble drugs. *ADMET and DMPK* **2020**, *8*, doi:10.5599/admet.844.

- 768 100. Nasrollahzadeh, M.; Sajadi, S.M.; Sajjadi, M.; Issaabadi, Z. Chapter 1 - An Introduction to  
769 Nanotechnology. In *Interface Science and Technology*; Nasrollahzadeh, M., Sajadi, S.M., Sajjadi,  
770 M., Issaabadi, Z., Atarod, M., Eds.; An Introduction to Green Nanotechnology; Elsevier, 2019;  
771 Vol. 28, pp. 1–27.
- 772 101. Kumar, P.S.; Pavithra, K.G.; Naushad, Mu. Chapter 4 - Characterization techniques for  
773 nanomaterials. In *Nanomaterials for Solar Cell Applications*; Thomas, S., Sakho, E.H.M.,  
774 Kalarikkal, N., Oluwafemi, S.O., Wu, J., Eds.; Elsevier, 2019; pp. 97–124 ISBN  
775 978-0-12-813337-8.
- 776 102. Mitić, Ž.; Stolić, A.; Stojanović, S.; Najman, S.; Ignjatović, N.; Nikolić, G.; Trajanović, M.  
777 Instrumental methods and techniques for structural and physicochemical characterization of  
778 biomaterials and bone tissue: A review. *Materials Science and Engineering: C* **2017**, *79*, 930–949,  
779 doi:10.1016/j.msec.2017.05.127.
- 780 103. Sosna-Głębska, A.; Sibiński, M.; Szczecińska, N.; Apostoluk, A. UV–Visible silicon detectors  
781 with zinc oxide nanoparticles acting as wavelength shifters. *Materials Today: Proceedings* **2020**,  
782 *20*, 25–29, doi:10.1016/j.matpr.2019.08.157.
- 783 104. Castillo, L.; Vargas, Z. Combined use of DSC, TGA, XDR and NIR in the compatibility study of  
784 preformulation mixtures for the development of 10 mg tablets of Rupatadine Fumarate. *Journal*  
785 *of Drug Delivery and Therapeutics* **2018**, *8*, 42–54, doi:10.22270/jddt.v8i3.1727.
- 786 105. Khanmohammadi, M.; Garmarudi, A.B.; Khoddami, N.; Shabani, K.; Khanlari, M. A novel  
787 technique based on diffuse reflectance near-infrared spectrometry and back-propagation  
788 artificial neural network for estimation of particle size in TiO<sub>2</sub> nano particle samples.  
789 *Microchemical Journal* **2010**, *95*, 337–340, doi:10.1016/j.microc.2010.01.020.
- 790 106. Riding, M.J.; Martin, F.L.; Trevisan, J.; Llabjani, V.; Patel, I.I.; Jones, K.C.; Semple, K.T.  
791 Concentration-dependent effects of carbon nanoparticles in gram-negative bacteria  
792 determined by infrared spectroscopy with multivariate analysis. *Environmental Pollution* **2012**,  
793 *163*, 226–234, doi:10.1016/j.envpol.2011.12.027.
- 794 107. Mitić, Ž.J.; Najman, S.J.; Cakić, M.D.; Ajduković, Z.R.; Ignjatović, N.L.; Nikolić, R.S.; Nikolić,  
795 G.M.; Stojanović, S.T.; Vukelić, M.Đ.; Trajanović, M.D. Spectroscopic characterization of bone  
796 tissue of experimental animals after glucocorticoid treatment and recovery period. *Journal of*  
797 *Molecular Structure* **2014**, *1074*, 315–320, doi:10.1016/j.molstruc.2014.06.006.
- 798 108. Boldon, L.; Laliberte, F.; Liu, L. Review of the fundamental theories behind small angle X-ray  
799 scattering, molecular dynamics simulations, and relevant integrated application. *Nano Reviews*  
800 **2015**, *6*, 25661, doi:10.3402/nano.v6.25661.
- 801 109. Schlögl, R. Chapter 5 X-ray Diffraction: A Basic Tool for Characterization of Solid Catalysts in  
802 the Working State. In *Advances in Catalysis*; Academic Press, 2009; Vol. 52, pp. 273–338.
- 803 110. Rajakumar, G.; Abdul, R. Larvicidal activity of synthesized silver nanoparticles using *Eclipta*  
804 *prostrata* leaf extract against filariasis and malaria vectors. *Acta Trop* **2011**, *118*, 196–203,  
805 doi:10.1016/j.actatropica.2011.03.003.
- 806 111. Farah, J.S.; Silva, M.C.; Cruz, A.G.; Calado, V. Differential calorimetry scanning: current  
807 background and application in authenticity of dairy products. *Current Opinion in Food Science*  
808 **2018**, *22*, 88–94, doi:10.1016/j.cofs.2018.02.006.

- 809 112. Ziegler-Borowska, M.; Chełminiak, D.; Kaczmarek, H.; Kaczmarek-Kędziera, A. Effect of side  
810 substituents on thermal stability of the modified chitosan and its nanocomposites with  
811 magnetite. *J Therm Anal Calorim* **2016**, *124*, 1267–1280, doi:10.1007/s10973-016-5260-x.
- 812 113. Valapa, R.; Pugazhenthii, G.; Katiyar, V. Thermal degradation kinetics of sucrose palmitate  
813 reinforced poly(lactic acid) biocomposites. *International Journal of Biological Macromolecules* **2014**,  
814 *65*, 275–283, doi:10.1016/j.ijbiomac.2014.01.053.
- 815 114. Mansfield, E.; Tyner, K.M.; Poling, C.M.; Blacklock, J.L. Determination of Nanoparticle Surface  
816 Coatings and Nanoparticle Purity Using Microscale Thermogravimetric Analysis. *Anal. Chem.*  
817 **2014**, *86*, 1478–1484, doi:10.1021/ac402888v.
- 818 115. Kwecińska, B.; Pusz, S.; Valentine, B.J. Application of electron microscopy TEM and SEM for  
819 analysis of coals, organic-rich shales and carbonaceous matter. *International Journal of Coal*  
820 *Geology* **2019**, *211*, 103203, doi:10.1016/j.coal.2019.05.010.
- 821 116. Falsafi, S.R.; Rostamabadi, H.; Assadpour, E.; Jafari, S.M. Morphology and microstructural  
822 analysis of bioactive-loaded micro/nanocarriers via microscopy techniques;  
823 CLSM/SEM/TEM/AFM. *Advances in Colloid and Interface Science* **2020**, *280*, 102166,  
824 doi:10.1016/j.cis.2020.102166.
- 825 117. Inkson, B.J. 2 - Scanning electron microscopy (SEM) and transmission electron microscopy  
826 (TEM) for materials characterization. In *Materials Characterization Using Nondestructive*  
827 *Evaluation (NDE) Methods*; Hübschen, G., Altpeter, I., Tschuncky, R., Herrmann, H.-G., Eds.;  
828 Woodhead Publishing, 2016; pp. 17–43 ISBN 978-0-08-100040-3.
- 829 118. Zuo, J.M.; Spence, J.C.H. Structure of Nanocrystals, Nanoparticles, and Nanotubes. In  
830 *Advanced Transmission Electron Microscopy: Imaging and Diffraction in Nanoscience*; Zuo, J.M.,  
831 Spence, J.C.H., Eds.; Springer: New York, NY, 2017; pp. 581–652 ISBN 978-1-4939-6607-3.
- 832 119. Gopinath, V.; MubarakAli, D.; Priyadarshini, S.; Priyadharshini, N.M.; Thajuddin, N.;  
833 Velusamy, P. Biosynthesis of silver nanoparticles from *Tribulus terrestris* and its antimicrobial  
834 activity: A novel biological approach. *Colloids and Surfaces B: Biointerfaces* **2012**, *96*, 69–74,  
835 doi:10.1016/j.colsurfb.2012.03.023.
- 836 120. Kumar, D.A.; Palanichamy, V.; Roopan, S.M. Green synthesis of silver nanoparticles using  
837 *Alternanthera dentata* leaf extract at room temperature and their antimicrobial activity.  
838 *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy* **2014**, *127*, 168–171,  
839 doi:10.1016/j.saa.2014.02.058.
- 840 121. Nakkala, J.R.; Mata, R.; Gupta, A.K.; Sadras, S.R. Biological activities of green silver  
841 nanoparticles synthesized with Acorous calamus rhizome extract. *European Journal of Medicinal*  
842 *Chemistry* **2014**, *85*, 784–794, doi:10.1016/j.ejmech.2014.08.024.
- 843 122. Nakkala, J.R.; Mata, R.; Bhagat, E.; Sadras, S.R. Green synthesis of silver and gold nanoparticles  
844 from *Gymnema sylvestre* leaf extract: study of antioxidant and anticancer activities. *J Nanopart*  
845 *Res* **2015**, *17*, 151, doi:10.1007/s11051-015-2957-x.
- 846 123. Katta, V.K.M.; Dubey, R.S. Green synthesis of silver nanoparticles using *Tagetes erecta* plant  
847 and investigation of their structural, optical, chemical and morphological properties. *Materials*  
848 *Today: Proceedings* **2020**, doi:10.1016/j.matpr.2020.02.809.
- 849 124. Zhang, X.-F.; Liu, Z.-G.; Shen, W.; Gurunathan, S. Silver Nanoparticles: Synthesis,  
850 Characterization, Properties, Applications, and Therapeutic Approaches. *International Journal*  
851 *of Molecular Sciences* **2016**, *17*, 1534, doi:10.3390/ijms17091534.

- 852 125. Mourdikoudis, S.; Pallares, R.; Thanh, N.T. Characterization techniques for  
853 nanoparticles: comparison and complementarity upon studying nanoparticle properties.  
854 *Nanoscale* **2018**, *10*, 12871–12934, doi:10.1039/C8NR02278J.
- 855 126. Zhang, L.; Mazouzi, Y.; Salmain, M.; Liedberg, B.; Boujday, S. Antibody-Gold Nanoparticle  
856 Bioconjugates for Biosensors: Synthesis, Characterization and Selected Applications. *Biosensors*  
857 *and Bioelectronics* **2020**, 112370, doi:10.1016/j.bios.2020.112370.
- 858 127. Cox, A.; Venkatachalam, P.; Sahi, S.; Sharma, N. Reprint of: Silver and titanium dioxide  
859 nanoparticle toxicity in plants: A review of current research. *Plant Physiol Biochem* **2016**, *110*,  
860 33–49, doi:10.1016/j.plaphy.2016.08.007.
- 861 128. Dasgupta, N.; Ranjan, S.; Mundekkad, D.; Ramalingam, C.; Shanker, R.; Kumar, A.  
862 Nanotechnology in agro-food: From field to plate. *Food Research International* **2015**, *69*, 381–400,  
863 doi:10.1016/j.foodres.2015.01.005.
- 864 129. Awad, M.A.; Eisa, N.E.; Virk, Promy.; Hendi, A.A.; Ortashi, K.M.O.O.; Mahgoub, A.S.A.;  
865 Elobeid, M.A.; Eissa, F.Z. Green synthesis of gold nanoparticles: Preparation, characterization,  
866 cytotoxicity, and anti-bacterial activities. *Materials Letters* **2019**, *256*, 126608,  
867 doi:10.1016/j.matlet.2019.126608.
- 868 130. Takeuchi, M.T.; Kojima, M.; Luetzow, M. State of the art on the initiatives and activities  
869 relevant to risk assessment and risk management of nanotechnologies in the food and  
870 agriculture sectors. *Food Research International* **2014**, *64*, 976–981,  
871 doi:10.1016/j.foodres.2014.03.022.
- 872 131. Marchiol, L.; Mattiello, A.; Pošćić, F.; Giordano, C.; Musetti, R. In vivo synthesis of  
873 nanomaterials in plants: location of silver nanoparticles and plant metabolism. *Nanoscale*  
874 *Research Letters* **2014**, *9*, 101, doi:10.1186/1556-276X-9-101.
- 875 132. Ribeiro, C.A.S.; Albuquerque, L.J.C.; de Castro, C.E.; Batista, B.L.; de Souza, A.L.M.;  
876 Albuquerque, B.L.; Zilse, M.S.; Bellettini, I.C.; Giacomelli, F.C. One-pot synthesis of  
877 sugar-decorated gold nanoparticles with reduced cytotoxicity and enhanced cellular uptake.  
878 *Colloids and Surfaces A: Physicochemical and Engineering Aspects* **2019**, *580*, 123690,  
879 doi:10.1016/j.colsurfa.2019.123690.
- 880 133. Vijayaraghavan, K.; Ashokkumar, T. Plant-mediated biosynthesis of metallic nanoparticles: A  
881 review of literature, factors affecting synthesis, characterization techniques and applications.  
882 *Journal of Environmental Chemical Engineering* **2017**, *5*, 4866–4883, doi:10.1016/j.jece.2017.09.026.
- 883 134. Chavan, R.R.; Bhinge, S.D.; Bhutkar, M.A.; Randive, D.S.; Wadkar, G.H.; Todkar, S.S.; Urade,  
884 M.N. Characterization, antioxidant, antimicrobial and cytotoxic activities of green synthesized  
885 silver and iron nanoparticles using alcoholic *Blumea eriantha* DC plant extract. *Materials Today*  
886 *Communications* **2020**, *24*, 101320, doi:10.1016/j.mtcomm.2020.101320.
- 887 135. El barghouti, M.; Akjouj, A.; Mir, A. Design of silver nanoparticles with graphene coatings  
888 layers used for LSPR biosensor applications. *Vacuum* **2020**, 109497,  
889 doi:10.1016/j.vacuum.2020.109497.
- 890 136. Teodoro, K.B.R.; Shimizu, F.M.; Scagion, V.P.; Correa, D.S. Ternary nanocomposites based on  
891 cellulose nanowhiskers, silver nanoparticles and electrospun nanofibers: Use in an electronic  
892 tongue for heavy metal detection. *Sensors and Actuators B: Chemical* **2019**, *290*, 387–395,  
893 doi:10.1016/j.snb.2019.03.125.

- 894 137. Giljohann, D.A.; Seferos, D.S.; Daniel, W.L.; Massich, M.D.; Patel, P.C.; Mirkin, C.A. Gold  
895 Nanoparticles for Biology and Medicine. *Angewandte Chemie International Edition* **2010**, *49*,  
896 3280–3294, doi:10.1002/anie.200904359.
- 897 138. Murphy, C.; Gole, A.; Stone, J.; Sisco, P.; Alkilany, A.; Goldsmith, E.; Baxter, S. Gold  
898 Nanoparticles in Biology: Beyond Toxicity to Cellular Imaging. *Accounts of chemical research*  
899 **2008**, *41*, 12, 1721–1730, doi:10.1021/ar800035u.
- 900 139. Bhattacharya, R.; Mukherjee, P. Biological properties of “naked” metal nanoparticles. *Advanced*  
901 *Drug Delivery Reviews* **2008**, *60*, 1289–1306, doi:10.1016/j.addr.2008.03.013.
- 902 140. Pardhi, D.M.; Şen Karaman, D.; Timonen, J.; Wu, W.; Zhang, Q.; Satija, S.; Mehta, M.; Charbe,  
903 N.; McCarron, P.A.; Tambuwala, M.; et al. Anti-bacterial activity of inorganic nanomaterials  
904 and their antimicrobial peptide conjugates against resistant and non-resistant pathogens.  
905 *International Journal of Pharmaceutics* **2020**, *586*, 119531, doi:10.1016/j.ijpharm.2020.119531.
- 906 141. Islam, N.U.; Jalil, K.; Shahid, M.; Rauf, A.; Muhammad, N.; Khan, A.; Shah, M.R.; Khan, M.A.  
907 Green synthesis and biological activities of gold nanoparticles functionalized with *Salix alba*.  
908 *Arabian Journal of Chemistry* **2019**, *12*, 2914–2925, doi:10.1016/j.arabjc.2015.06.025.
- 909 142. Rashmi, B.N.; Harlapur, S.F.; Avinash, B.; Ravikumar, C.R.; Nagaswarupa, H.P.; Anil Kumar,  
910 M.R.; Gurushantha, K.; Santosh, M.S. Facile green synthesis of silver oxide nanoparticles and  
911 their electrochemical, photocatalytic and biological studies. *Inorganic Chemistry Communications*  
912 **2020**, *111*, 107580, doi:10.1016/j.inoche.2019.107580.
- 913 143. Yugay, Y.A.; Usoltseva, R.V.; Silant'ev, V.E.; Egorova, A.E.; Karabtsov, A.A.; Kumeiko, V.V.;  
914 Ermakova, S.P.; Bulgakov, V.P.; Shkryl, Y.N. Synthesis of bioactive silver nanoparticles using  
915 alginate, fucoidan and laminaran from brown algae as a reducing and stabilizing agent.  
916 *Carbohydrate Polymers* **2020**, *245*, 116547, doi:10.1016/j.carbpol.2020.116547.
- 917 144. Ramesh, A.; Tamizhdurai, P.; Gopinath, S.; Sureshkumar, K.; Murugan, E.; Shanthi, K. Facile  
918 synthesis of core-shell nanocomposites Au catalysts towards abatement of environmental  
919 pollutant Rhodamine B. *Heliyon* **2019**, *5*, e01005, doi:10.1016/j.heliyon.2018.e01005.
- 920 145. Thangamani, N.; Bhuvaneshwari, N. Green synthesis of gold nanoparticles using *Simarouba*  
921 *glauca* leaf extract and their biological activity of micro-organism. *Chemical Physics Letters* **2019**,  
922 *732*, 136587, doi:10.1016/j.cplett.2019.07.015.
- 923 146. Lediga, M.E.; Malatjie, T.S.; Olivier, D.K.; Ndinteh, D.T.; van Vuuren, S.F. Biosynthesis and  
924 characterisation of antimicrobial silver nanoparticles from a selection of fever-reducing  
925 medicinal plants of South Africa. *South African Journal of Botany* **2018**, *119*, 172–180,  
926 doi:10.1016/j.sajb.2018.08.022.
- 927 147. Montes de Oca-Vásquez, G.; Solano-Campos, F.; Vega-Baudrit, J.R.; López-Mondéjar, R.;  
928 Odriozola, I.; Vera, A.; Moreno, J.L.; Bastida, F. Environmentally relevant concentrations of  
929 silver nanoparticles diminish soil microbial biomass but do not alter enzyme activities or  
930 microbial diversity. *Journal of Hazardous Materials* **2020**, *391*, 122224,  
931 doi:10.1016/j.jhazmat.2020.122224.
- 932 148. Nie, X.; Zhu, K.; Zhao, S.; Dai, Y.; Tian, H.; Sharma, V.K.; Jia, H. Interaction of Ag<sup>+</sup> with soil  
933 organic matter: Elucidating the formation of silver nanoparticles. *Chemosphere* **2020**, *243*,  
934 125413, doi:10.1016/j.chemosphere.2019.125413.

- 935 149. Hashimoto, Y.; Takeuchi, S.; Mitsunobu, S.; Ok, Y.-S. Chemical speciation of silver (Ag) in soils  
936 under aerobic and anaerobic conditions: Ag nanoparticles vs. ionic Ag. *J. Hazard. Mater.* **2017**,  
937 322, 318–324, doi:10.1016/j.jhazmat.2015.09.001.
- 938 150. Mirzajani, F.; Askari, H.; Hamzelou, S.; Schober, Y.; Römpf, A.; Ghassempour, A.; Spengler, B.  
939 Proteomics study of silver nanoparticles toxicity on *Oryza sativa* L. *Ecotoxicology and*  
940 *Environmental Safety* **2014**, *108*, 335–339, doi:10.1016/j.ecoenv.2014.07.013.
- 941 151. Qian, H.; Peng, X.; Han, X.; Ren, J.; Sun, L.; Fu, Z. Comparison of the toxicity of silver  
942 nanoparticles and silver ions on the growth of terrestrial plant model *Arabidopsis thaliana*.  
943 *Journal of Environmental Sciences* **2013**, *25*, 1947–1956, doi:10.1016/S1001-0742(12)60301-5.
- 944 152. Cox, A.; Venkatachalam, P.; Sahi, S.; Sharma, N. Silver and titanium dioxide nanoparticle  
945 toxicity in plants: A review of current research. *Plant Physiology and Biochemistry* **2016**, *107*, 147–  
946 163, doi:10.1016/j.plaphy.2016.05.022.
- 947 153. El-Temsah, Y.S.; Joner, E.J. Impact of Fe and Ag nanoparticles on seed germination and  
948 differences in bioavailability during exposure in aqueous suspension and soil. *Environmental*  
949 *Toxicology* **2012**, *27*, 42–49, doi:10.1002/tox.20610.
- 950 154. Kaveh, R.; Li, Y.-S.; Ranjbar, S.; Tehrani, R.; Brueck, C.L.; Van Aken, B. Changes in *Arabidopsis*  
951 *thaliana* Gene Expression in Response to Silver Nanoparticles and Silver Ions. *Environ. Sci.*  
952 *Technol.* **2013**, *47*, 10637–10644, doi:10.1021/es402209w.
- 953 155. Taylor, A.F.; Rylott, E.L.; Anderson, C.W.N.; Bruce, N.C. Investigating the Toxicity, Uptake,  
954 Nanoparticle Formation and Genetic Response of Plants to Gold. *PLOS ONE* **2014**, *9*, e93793,  
955 doi:10.1371/journal.pone.0093793.
- 956 156. Courtois, P.; Rorat, A.; Lemiere, S.; Guyoneaud, R.; Attard, E.; Levard, C.; Vandebulcke, F.  
957 Ecotoxicology of silver nanoparticles and their derivatives introduced in soil with or without  
958 sewage sludge: A review of effects on microorganisms, plants and animals. *Environmental*  
959 *Pollution* **2019**, *253*, 578–598, doi:10.1016/j.envpol.2019.07.053.
- 960 157. Mishra, S.; Yang, X.; Singh, H.B. Evidence for positive response of soil bacterial community  
961 structure and functions to biosynthesized silver nanoparticles: An approach to conquer  
962 nanotoxicity? *Journal of Environmental Management* **2020**, *253*, 109584,  
963 doi:10.1016/j.jenvman.2019.109584.
- 964 158. Eivazi, F.; Afrasiabi, Z.; Jose, E. Effects of Silver Nanoparticles on the Activities of Soil Enzymes  
965 Involved in Carbon and Nutrient Cycling. *Pedosphere* **2018**, *28*, 209–214,  
966 doi:10.1016/S1002-0160(18)60019-0.
- 967 159. Hänsch, M.; Emmerling, C. Effects of silver nanoparticles on the microbiota and enzyme  
968 activity in soil. *Journal of Plant Nutrition and Soil Science* **2010**, *173*, 554–558,  
969 doi:10.1002/jpln.200900358.
- 970 160. Shin, Y.-J.; Kwak, J.I.; An, Y.-J. Evidence for the inhibitory effects of silver nanoparticles on the  
971 activities of soil exoenzymes. *Chemosphere* **2012**, *88*, 524–529,  
972 doi:10.1016/j.chemosphere.2012.03.010.
- 973 161. Peyrot, C.; Wilkinson, K.J.; Desrosiers, M.; Sauvé, S. Effects of silver nanoparticles on soil  
974 enzyme activities with and without added organic matter. *Environmental Toxicology and*  
975 *Chemistry* **2014**, *33*, 115–125, doi:10.1002/etc.2398.



- 976 162. González-Fuenzalida, R.A.; Sanjuan-Navarro, L.; Moliner-Martínez, Y.; Campíns-Falcó, P.  
977 Quantitative study of the capture of silver nanoparticles by several kinds of soils. *Science of The*  
978 *Total Environment* **2018**, *630*, 1226–1236, doi:10.1016/j.scitotenv.2018.02.307.
- 979 163. Grün, A.-L.; Straskraba, S.; Schulz, S.; Schloter, M.; Emmerling, C. Long-term effects of  
980 environmentally relevant concentrations of silver nanoparticles on microbial biomass, enzyme  
981 activity, and functional genes involved in the nitrogen cycle of loamy soil. *Journal of*  
982 *Environmental Sciences* **2018**, *69*, 12–22, doi:10.1016/j.jes.2018.04.013.
- 983 164. Rahmatpour, S.; Shirvani, M.; Mosaddeghi, M.R.; Nourbakhsh, F.; Bazarganipour, M. Dose-  
984 response effects of silver nanoparticles and silver nitrate on microbial and enzyme activities in  
985 calcareous soils. *Geoderma* **2017**, *285*, 313–322, doi:10.1016/j.geoderma.2016.10.006.
- 986 165. McGee, C.F.; Storey, S.; Clipson, N.; Doyle, E. Soil microbial community responses to  
987 contamination with silver, aluminium oxide and silicon dioxide nanoparticles. *Ecotoxicology*  
988 **2017**, *26*, 449–458, doi:10.1007/s10646-017-1776-5.
- 989 166. Asadishad, B.; Chahal, S.; Akbari, A.; Cianciarelli, V.; Azodi, M.; Ghoshal, S.; Tufenkji, N.  
990 Amendment of Agricultural Soil with Metal Nanoparticles: Effects on Soil Enzyme Activity  
991 and Microbial Community Composition. *Environ. Sci. Technol.* **2018**, *52*, 1908–1918,  
992 doi:10.1021/acs.est.7b05389.
- 993 167. Grün, A.-L.; Manz, W.; Kohl, Y.L.; Meier, F.; Straskraba, S.; Jost, C.; Drexel, R.; Emmerling, C.  
994 Impact of silver nanoparticles (AgNP) on soil microbial community depending on  
995 functionalization, concentration, exposure time, and soil texture. *Environ Sci Eur* **2019**, *31*, 15,  
996 doi:10.1186/s12302-019-0196-y.
- 997 168. Meier, M.J.; Dodge, A.E.; Samarajeewa, A.D.; Beaudette, L.A. Soil exposed to silver  
998 nanoparticles reveals significant changes in community structure and altered microbial  
999 transcriptional profiles. *Environmental Pollution* **2020**, *258*, 113816,  
1000 doi:10.1016/j.envpol.2019.113816.
- 1001 169. Li, M.; Greenfield, B.K.; Nunes, L.M.; Dang, F.; Liu, H.; Zhou, D.; Yin, B. High retention of  
1002 silver sulfide nanoparticles in natural soils. *Journal of Hazardous Materials* **2019**, *378*, 120735,  
1003 doi:10.1016/j.jhazmat.2019.06.012.