1 Trophic guild and forest type explain phyllostomid bat abundance variation from

2 human habitat disturbance.

- 3 Jorge D. Carballo-Morales¹, Romeo A. Saldaña-Vázquez^{2,3*}, Federico Villalobos¹
- 4 1. Laboratorio de Sistemática, Genética y Evolución, Escuela de Ciencias Biológicas,
- 5 Universidad Nacional, Heredia, CP 3000, Costa Rica.
- 6 2. Instituto de Investigaciones en Medio Ambiente Xavier Gorostiaga, S.J. Universidad
- 7 Iberoamericana Puebla, Blvd. del Niño Poblano No. 2901, Col. Reserva Territorial Atlixcáyotl,
- 8 San Andrés Cholula, Puebla C. P. 72820.
- 9 3. Facultad de Ciencias Biológicas, Benemérita Universidad Autónoma de Puebla. Blvd.,
- 10 Valsequillo y Av., San Claudio, Edificio BIO 1, Ciudad Universitaria, Col. Jardines de San
- 11 Manuel, C.P. 72570 Puebla, México.
- 12 *Corresponding author:
- 13 E-mail address: romeo.saldana@gmail.com (R. A. Saldaña-Vázquez)

14 Trophic guild and forest type explain phyllostomid bat abundance variation from

15 human habitat disturbance.

16 Highlights

- 17 I No phylogenetic signal was found in the abundance of bats in disturbed habitats.
- 18 The abundance of bats in disturbed habitats depends on the trophic guild.
- 19 Porest altitude influences the abundance of bats in disturbed habitats.
- 20 Animal-feeding bats reduced their abundance in disturbed habitats regardless of
 21 disturbance type and forest type.

22 Abstract

23 The loss of tropical forest cover caused by land-use change is causing a reduction in 24 functional groups, such as trophic guilds. Phyllostomid bats (family Phyllostomidae) are 25 essential in the Neotropics since they occupy up to six trophic guilds, and are pollinators. 26 seed dispersers, and regulators of vertebrate and invertebrate populations. In this study, a 27 series of meta-analyses were performed in order to analyse their response to habitat 28 disturbance. Data were obtained through a comprehensive literature review whereby we 29 measured the abundance of phyllostomid bats in disturbed habitats and conserved forests. 30 We found that the abundance of phytophagous bats depends on the type of habitat 31 disturbance and the type of forest where it occurs. On the other hand, animal-feeding bats 32 reduce their abundance in any disturbed habitat regardless of disturbance type and forest 33 type. No phylogenetic signal was found in the response of bats to habitat disturbance, nor 34 was the response found to be dependent on the type of crop, the age of the secondary forest, 35 or the distance to a conserved forest. These results demonstrate that feeding and the type of

- 36 forest where the disturbance occurs are important aspects to understand the reduction of
- 37 animal populations in the face of habitat destruction processes. This has implications on the
- 38 conservation of species and their function in ecosystems.

39 Keywords

Chiroptera, Neotropical leaf-nosed bats, land-use change, phylogenetic signal, meta-analysis,
distance to conserved forest.

42 **1. Introduction**

43 Tropical ecosystems are losing their forest cover due to changes in land use resulting from 44 the development of different human activities (Venter et al. 2016; Potapov et al. 2017). 45 Urbanization, logging, agriculture, and livestock are the main land uses responsible for the 46 current deforestation (Potapov et al. 2017). In many cases, these can cause a reduction in the abundance of species (Dirzo et al. 2014; Newbold et al. 2015) and the subsequent loss of 47 48 ecosystem functions due the reduction in functional groups. For example, a reduction in 49 trophic guilds that fulfil specific functions, such as pollination, seed dispersal, and control of 50 populations within ecosystems, results in an imbalance of ecosystem functionality (Díaz et al. 51 2013; Dirzo et al. 2014; Newbold et al. 2020).

In the Neotropics, leaf-nosed bats (Phyllostomidae) are essential in ecosystem
functionality. They are pollinators and seed dispersers of more than 700 plants species, and
they are also predators of vertebrates and invertebrates, thus acting as regulators of their
populations (Muscarella & Fleming 2007; Kalka et al. 2008; Kunz et al. 2011). The family

56 Phyllostomidae consists of 11 subfamilies, 60 genera, and 212 species (Cirranello et al. 2016) 57 distributed throughout the tropics and subtropics of America, including the Antilles (Villalobos 58 & Arita 2010). Species diversity patterns of phyllostomids are considered one of the greatest 59 adaptive radiations among vertebrate families due to the wide ecological and morphological 60 variation they exhibit (Freeman 2000; Dumont et al. 2014). This morphological variation is 61 reflected in six trophic guilds: nectarivores, frugivores, insectivores, carnivores, sanguivores, 62 and omnivores (Freeman 2000; Rex et al. 2010; Rojas et al. 2011). In addition, phyllostomids 63 have been considered good bioindicators in Neotropical forests, since their species richness 64 and relative abundance is reduced by human habitat disturbance (Fenton et al. 1992;

65 Medellín et al. 2000; Jones et al. 2009).

66 Previous studies suggest that the reduction in the populations of phyllostomid species 67 depends on the trophic guild to which they belong because not all tropics guilds are affected 68 in the same way by habitat disturbance (Medellín et al. 2000; Klingbeil & Willig 2009; Willig et 69 al. 2019). In general, frugivorous and nectarivorous bats are considered tolerant to habitat 70 disturbance, whereas insectivorous and carnivorous bats are considered sensitive. However, 71 when analysing the response of bats from different trophic guilds in different types of habitat 72 disturbance, contradictory results arise. This contradictory pattern has been observed in 73 nectarivorous (Ochoa 2000; Williams-Guillén & Perfecto 2010; Durán & Pérez 2015), 74 insectivorous (Murillo-García & Bedoya-Durán 2014), and sanguivorous phyllostomid bats 75 (Quinto-Mosquera et al. 2013; Gonçalves et al. 2017). The latter, traditionally considered as 76 tolerant to human disturbance activities, such as livestock, appear to be sensitive (Quinto-77 Mosquera et al. 2013; Gonçalves et al. 2017).

78 Several studies have been conducted in order to explore this controversy. It has been 79 found that omnivorous and frugivorous bats are tolerant to livestock grassland, whereas 80 carnivorous, insectivorous, nectarivorous, and sanguivorous bats become sensitive to this 81 type of disturbance (Goncalves et al. 2017). Another study that considered different types of 82 habitat disturbance showed that carnivorous and insectivorous bats are sensitive to habitat 83 disturbance, frugivores and nectarivores tolerate agroforestry crops, and all trophic guilds, 84 including omnivores and sanguivores, are sensitive to monocultures and grasslands (García-85 Morales et al. 2013). This is consistent with the results of a recent study that found that the 86 functional and taxonomic diversity of Neotropical bats decreases in habitats less similar to 87 conserved forests (i.e. high contrast), such as crops, grasslands, and early-stage secondary 88 forests (Farneda et al. 2019). However, it is not clear how bats belonging to different trophic 89 guilds would respond to urbanization, logging, and tourism. In addition, considering the wide 90 aeographic distribution of phyllostomids, it is still unknown if the type of forest where the 91 disturbance occurs could affect their relative abundance. Moreover, there are factors that 92 have not been considered, such as the distance between conserved forests and disturbed 93 habitats, and the phylogenetic signal in the response to disturbance. Previous studies have 94 shown that the abundance of frugivorous bats decreases in wooded crops and grasslands as 95 the distance to a patch of conserved forest increases (Galindo-González & Sosa 2003), which 96 is similar to what has been found in birds and trees (Socolar et al. 2019). It is important to 97 note that, because we are performing observations with bat species, it is necessary to 98 determine the phylogenetic signal based on the phylogenetic relationship between species. 99 since their relatedness could generate statistical non-independence (Münkemüller et al.

2012). Finally, since the last meta-analysis carried out in 2013, more than twenty studies have
been conducted in the Neotropics in order to understand the response of phyllostomids to
habitat disturbance. These studies can allow to make a new quantitative revision about the
effect of habitat disturbance on phyllostomid abundance.

104 Therefore, the objective of this study was to answer, through a series of meta-

105 analyses, the following questions:

106 i) Is the relative abundance of trophic guilds occupied by phyllostomids different 107 according to the type of habitat disturbance? We expected frugivorous, nectarivorous, and 108 omnivorous bats to be more abundant, or maintain a similar abundance, in disturbed forests, 109 such as secondary forests, crops, mixed habitats with crops and grasslands, forests with 110 selective logging, forests with tourism, and urban areas, compared to conserved forests. 111 These types of human disturbances allow the presence of pioneer plants in these disturbed 112 habitats or their surroundings, favouring phytophagous bats or those that include pioneer 113 plants in their diet (Peters et al. 2006; Castro-Luna et al. 2007; Saldaña-Vázquez et al. 2010; 114 Prone et al. 2012; Cisneros et al. 2015; Willig et al. 2019). We expected sanguivores to 115 increase their abundance, or maintain a similar abundance, in secondary forests, crops, and 116 mixed habitats with crops and grasslands, compared to conserved forests. The proximity of 117 these habitats to human settlements and their domestic animals represents a potential source 118 of food (Delpietro et al. 1992; Bobrowiec et al. 2015). Conversely, we expected carnivores 119 and insectivores to be more abundant in conserved forests, compared to all other types of 120 habitat disturbance, since deforestation promotes a reduction in prey and roosting sites 121 (Bernard & Fenton 2003; Jones et al. 2017).

122 ii) Is the relative abundance of trophic guilds of phyllostomids dependent on crop 123 intensity and secondary forest age? We expected carnivores and insectivores to be sensitive 124 regardless of crop intensity or secondary forest age, based on our previous predictions. In the 125 case of sanguivores, we expected their response to not be related to crop intensity or forest 126 age, since it is believed that their response depends rather on the proximity to human 127 settlements, as mentioned above. We expected frugivores, nectarivores, and omnivores to be 128 tolerant to low intensity crops (e.g. agroforestry) and sensitive to high intensity crops (e.g. 129 monocultures), since pioneer plants have been observed to be abundant in low intensity 130 crops, and their abundance decreases as intensity increases (Williams-Guillén & Perfecto 131 2010). We also expected these three guilds to remain tolerant throughout the different ages of 132 secondary forests, since pioneer plants are abundant in these habitats due to the succession 133 process (Castro-Luna et al. 2007; de la Peña-Cuéllar et al. 2012; Farneda et al. 2018).

iii) Is the relative abundance of phyllostomid trophic guilds in disturbed habitats related
to the distance to conserved forests? We expected bats, despite their ability to fly, to
decrease their abundance in disturbed habitats as the distance to conserved forests
increased, which is similar to patterns observed in terrestrial mammals, other bats, and birds
(Galindo-González & Sosa 2003; Socolar et al. 2019; Pardo et al. 2019). We expected this
because most diurnal refuges of bats are in conserved habitats (Cortés-Delgado & Sosa
2014).

iv) Is the relative abundance of phyllostomid trophic guilds related to the forest where
human disturbance occurs? We expected bats to be sensitive to different types of habitat
disturbance in forests with higher elevation but not in lowland tropical forests, as has been

observed in another animal groups (Dalsgaard et al. 2018). This is because there is a greater
availability of food in lowland tropical forests (<1000 meters), which facilitates the increase in
bat populations and maintains a greater diversity of bat species compared to forests of higher
elevation (Rex et al. 2008; Martins et al. 2015).

148

149 **2. Methods**

150 2.1. Literature search

151 We conducted an extensive review of the available literature through Google scholar. The

152 keywords used were "phyllostomidae", "abundancia", "alteración", "perturbación",

153 "murciélagos", "abundance", "perturbation", and "bats". We did not include words in

154 Portuguese because most studies published in this language regularly include a title, abstract,

and keywords in English. We did not limit the search by year of publication. We selected

156 studies that sampled phyllostomids in disturbed habitats and at least one conserved forest. All

157 the studies considered in the analysis included: number of captures of each bat species per

158 site, distance between sampled sites, and a description of the type of habitat disturbance.

159 When studies did not report the distance among sampled sites, we extracted it from the study

160 site map (when present) with ImageJ 1.52a (Schneider et al. 2012).

161

162 2.2. Database

We obtained a total of 22 studies presenting useful data (Supporting Information). The 22 studies summarized 763 bat species observations (i.e. number of cases, k) conducted in six countries (Fig. 1). The observations comprised 107 phyllostomid bat species belonging to 42 genera and 11 trophic guilds, based on (Rojas et al. 2011), and seven types of disturbance (Table 1) and five forest types, according to the ecological zones defined by the FAO (2012).

168

169 2.3. Phylogenetic signal in phyllostomid abundance

170 We identified two potential sources of non-independence in our data. The first one was the 171 possibility of phylogenetic signal in the species abundance observed in disturbed habitats 172 (Nakagawa et al. 2017). To assess this effect, we evaluated the presence of phylogenetic 173 signal by making a phylogenetic tree of the species present in our database. The second 174 source was related to the author involved in the investigation (Nakagawa et al. 2017), when 175 more than one species data item coming from the same author could be similar or biased. To 176 deal with this effect, we included author as a random effect in our meta-analysis (see below). 177 For the estimation of phylogenetic signal in species abundance in disturbed habitats, we 178 constructed a new phylogeny for the Phyllostomidae species present in our abundance 179 dataset, since not all species with observations of abundance were present in previous 180 Phyllostomidae phylogenies (Baker & Hoofer 2003; Datzmann et al. 2010; Rojas et al. 2011); 181 Shi and Rabosky (2015)). Characters used were DNA sequences obtained from the Genbank 182 database (NCBI) for Cytochrome Oxidase I (COI), Recombination activating 2 (RAG2), and 183 Cytochrome B (CYTB) genes from 157 phyllostomid species and one mormoopid species

184 (Pteronotus parnellii), which was used as an outgroup (Supporting Information). We aligned 185 all sequences with MUSCLE using the default parameters (Edgar 2004). We performed a 186 multilocus phylogenetic reconstruction in BEAST v1.8.4 (Suchard et al. 2018) for the 187 Bayesian inference. We linked the trees and the molecular substitution model for all genes. 188 We used the GTR+I+G molecular substitution model, strict clock model, and constant size 189 model for the construction of the tree (Kingman 1982). We did three independent runs for one 190 hundred million generations and sampled every one thousand generations. We discarded the 191 first 2.5 million generations of each run as a burn-in. We used TRACER v1.8.2 (Rambaut et 192 al. 2018) to estimate the effective sample size for each parameter (all resulting effective 193 sample sizes exceeded 100) and its convergence, and to calculate the mean, upper, and 194 lower bounds of the 95% highest posterior density interval (95% HPD). We combined the 195 trees sampled from each independent run (10000) using LogCombiner and TreeAnnotator 196 (Heled & Drummond 2010). The obtained grouping of species was consistent with 197 phylogenetic trees generated in other studies (Rojas et al. 2011; Shi & Rabosky 2015). 198 We looked for a phylogenetic signal in the abundance of bat species by performing a randomization test in the R computational environment (R Core Team 2018). We used the 199

obtained phylogenetic tree (Fig. 2) and its branch lengths to perform the randomization test,
which evaluates the variation expected in a quantitative trait under a Brownian motion model
of evolution compared with values obtained by shuffling trait data across the tips. Higher
values and statistically significant values of the K index indicate a stronger phylogenetic signal
(Blomberg et al. 2003). Species abundance values used in the randomization test were the
mean of the proportion of bats captured in disturbed habitats for each species. To obtain

these values, we performed a multivariate meta-analysis of the proportion of the abundance
of species captured in disturbed habitats, taking species as a fixed factor and the author that
reported the abundance value as a random factor (Viechtbauer 2010).

209 Given that not all phyllostomid species with abundance were present in the 210 phylogenetic tree, we pruned the branches using the "Picante" package (Kembel et al. 2010). 211 In this way, species in the tree corresponded to those in the database used in the meta-212 analysis. Of the 158 species in the tree and 107 species in the database, only 91 were used 213 in the analysis (Supporting Information). We used the "Geiger" package (Harmon et al. 2008) 214 to create a file that contained the species, the effect size, and the phylogenetic lengths. The 215 phylogenetic signal was tested with the "phytools" package (Revell 2012) using 1000 216 randomizations. Due to the absence of phylogenetic signal in the proportion of bat abundance 217 in the assessed species (K=0.35, P= 0.204, Fig. 2), the meta-analyses below were performed 218 without phylogenetic correction.

219

220 2.4. Effect of habitat disturbance on abundance of phyllostomid trophic guilds

We performed five meta-analyses corresponding to each question in our objectives and their respective moderator variables: 1) habitat disturbance type, 2) crop type, 3) secondary forest age, 4) distance between conserved and disturbed sample sites, and 5) habitat disturbance type inside each forest type. The selected effect size was the proportion of individuals captured in the disturbed habitat from the total of individuals captured per species in both habitats (conserved and disturbed); hereafter referred to as "bat abundance". In each metaanalysis, the author of the study was considered as a random factor (Nakagawa et al. 2017).
All analyses were performed using the package "Metafor" (Viechtbauer 2010) in the R
computational environment (R Core Team 2018).

230 We used Cochran's Q index as a measure of heterogeneity for each analysis. 231 Heterogeneity is important in meta-analyses because it allows to evaluate if the variation in 232 the collected effect sizes is explained by population variation or by chance (Harrison 2011; 233 Nakagawa et al. 2017). In addition, if heterogeneity is significant, it means that variation in 234 effect size could be explained by moderator variables (i.e. forest type, distance, etc.). In order 235 to examine the publication bias in our data set, we performed a regression test (Egger et al. 236 1997; Nakagawa et al. 2017). A significant result in the regression test indicates that effect 237 sizes (i.e. bat abundance) are balanced. Finally, we did not perform a meta-analysis for 238 carnivorous (C) guilds due to the low number of observations in the database (k = 2).

239

240 **3. Results**

- 241 Bat abundance in disturbed habitats was 0.46 from the total abundance observed (P<0.0001,
- 242 CI=0.42-0.48, k=763), and the observations were significantly heterogeneous
- 243 (Q=28237.5540, d.f.=762, P<0.0001). Therefore, the response of bats to habitat disturbance
- was classified as sensitive because the confidence intervals (CI) of the abundance estimate
- in disturbed habitats was lower than 0.5 and, thus, did not overlap with this value.
- 246 In general, we found that frugivores (F), frugivores-nectarivores (FN), insectivores-
- 247 nectarivores (IN), insectivores-nectarivores-frugivores (INF), and nectarivores (N) were

tolerant to habitat disturbance. On the other hand, insectivores (I), sanguivores (S),

insectivores-frugivores (IF), carnivores-frugivores (CF), and insectivores-carnivores-frugivores
(ICF) were sensitive to habitat disturbance (Supporting Information). In addition, for these last
three trophic guilds (IF, CF and ICF), the heterogeneity of the adjusted models was not
significant (Supporting Information). Therefore, we did not evaluate the effect of the

253 moderator variables on the abundance of these guilds.

We evaluated the different moderator variables of bat abundance in disturbed habitats for bats from trophic guilds with significant heterogeneity (F, FN, IN, INF and N). We did not find differences in the abundance of these trophic guilds among different crop types (Supporting Information) despite the significant heterogeneity of the adjusted model (P <0.05, Supporting Information). Secondary forests age and distance from conserved forests were partly explained by trophic guild ($\square_{distance}=0.1-0.5$, Supporting Information); however, there was no significance in their heterogeneity (P>0.05, Supporting Information).

In the meta-analysis with habitat disturbance type as moderator, we found that only F, N, and INF responded significantly different (Table 2). We found that F were sensitive to logging, grassland, and urbanization, but tolerant to secondary forest and crops. Nectarivores were sensitive to urbanization and tolerant to secondary forest and crops. Finally, INF were sensitive to logging, grassland, and crops, but they were tolerant to secondary forest and mixed habitats such as grassland-crops.

267 Our nested analysis indicated that the response of bats depends on the forest type 268 where the disturbance occurs only in the case of F and FN (Table 3). Frugivores were

sensitive to secondary forest if it occurred in the Tropical Mountain System but tolerant when it occurred in the Tropical Rainforest. They were also sensitive to urbanization if it occurred in the Tropical Moist Forest or the Tropical Mountain System but tolerant when it occurred in the Tropical Rainforest. Frugivores-nectarivores were sensitive to secondary forest if it occurred in the Tropical Mountain System but tolerant when it occurred in the Tropical Dry Forest or Tropical Rainforest and were sensitive to crops if they occurred in the Tropical Mountain System but tolerant when they occurred in the Tropical Mountain

276

277 **4. Discussion**

278 Our results show that populations of phyllostomid bats are sensitive to human habitat 279 disturbance, which contrasts with what was found in a previous meta-analysis (García-280 Morales et al. 2013). However, we found that this response depends on the trophic guild to 281 which the species belong. We observed that frugivores (F), frugivores-nectarivores (FN), 282 insectivores-nectarivores (IN), insectivores-nectarivores-frugivores (INF), and nectarivores (N) 283 were tolerant to habitat disturbance, whereas insectivores (I), sanguivores (S), insectivores-284 frugivores (IF), carnivores-frugivores (CF), and insectivores-carnivores-frugivores (ICF) were 285 sensitive. These results agree with previous observations of the effect of habitat disturbance 286 on phyllostomid abundance (Fenton et al. 1992; Medellín et al. 2000; Ávila-Gómez et al. 287 2015).

Even though I and S showed significance in the heterogeneity of the adjusted models, they remained sensitive in subsequent analyses with moderator variables. The response to habitat disturbance by sensitive guilds could be explained by initial deforestation causing the
loss of potential refuges. For example, S use holes in living trees as well as caves or cracks in
rocks (Voss et al. 2016; Gonçalves et al. 2017), while I and C use holes in living trees or
arboreal termite nests (Bernard & Fenton 2003; Kalko et al. 2006; Jones et al. 2017). In
addition, I and C present fidelity and permanence towards this resource. Moreover, the prey
of these species are more abundant in conserved forests (Kalko et al. 1999; de la PeñaCuéllar et al. 2012).

297 4.1 Habitat disturbance type

Different types of habitat disturbance promote different changes in food and other resources used by phyllostomids and, thus, not all bats can tolerate disturbance in a similar way. We found F and N to have the highest variation in abundance response in relation to disturbance type. This pattern was previously observed by García-Morales et al. (2013).

302 We also found that F and N were tolerant to secondary forests and crops, which is 303 consistent with previous studies (Fenton et al. 1992; Klingbeil & Willig 2009). This response 304 may be due to the abundance of chiropterochorous or chiropterophilous species among 305 pioneer plants in these habitats, thus favouring F and N (Castro-Luna et al. 2007; Muscarella 306 & Fleming 2007; Castro-Luna & Galindo-González 2012). Frugivores were also tolerant to 307 forests with tourism; however, our results are not conclusive because we only included one 308 study with this type of habitat disturbance. This study was conducted near a conserved forest 309 and a secondary forest, which explains the high abundance of this trophic guild (Murillo-310 García & Bedoya-Durán 2014). On the other hand, we expected F and N to be tolerant to

311	urban zones and forests with selective logging, as has been observed in previous studies
312	(Peters et al. 2006; Ferreira et al. 2010; Prone et al. 2012). However, we found the opposite,
313	which could be explained by our analysis being performed at the trophic guild level, whereas
314	studies that found these guilds to be tolerant were performed at the genus level (Saldaña-
315	Vázquez et al. 2010; Saldaña-Vázquez & Schondube 2016). Finally, we found F to be
316	sensitive to livestock grasslands, which is consistent with our expectations and the study by
317	García-Morales et al. (2013).
318	
319	4.2. Crop type and secondary forest age
320	We found that crop type and secondary forest age did not affect the abundance of
321	phyllostomid trophic guilds. This could be explained by our study including a high number of
322	studies on agroforestry crops versus only one on monocultures. Similarly, in the case of
323	secondary forest age, most studies evaluated secondary forests of 15 years or less, whereas
324	very few looked at secondary forests of 50 years or more. The results may change with data
325	with the same number of cases per crop type according to intensity and secondary forests of

- 326 different ages.
- 327
- 328 4.3. Distance to conserved forests

We found that the distance to conserved forests does not influence the abundance ofphyllostomid trophic guilds in disturbed habitats. Other studies show the opposite effect in

non-flying mammals, bats, birds, and trees (Galindo-González & Sosa 2003; Cleary et al.

2016; Socolar et al. 2019; Pardo et al. 2019). The lack of a significant effect of distance to
conserved habitats on bat abundance in disturbed habitats may be related to the variation in
habitat disturbance type in our study and the ability of phyllostomids to fly large distances
either to migrate or forage (Arnone et al. 2016; Esbérard et al. 2017; Medellin et al. 2018). In
order to determine the effect of these two variables on bat abundance in disturbed habitats,
studies that evaluate the effects of habitat disturbance on bat abundance comparing
disturbance type and phyllsotomid vagility are necessary.

339

340 4.4. Forest type

Our results show that the abundance of F and FN can vary depending on the type of forest where the habitat disturbance occurs. Both trophic guilds decreased their abundance, from tolerant to sensitive, in secondary forests of tropical mountain systems (forests with altitudes higher than 1000 meters). The same happened with FN, which became sensitive in crops located in tropical mountain systems. On the other hand, F became tolerant in urban zones when they occurred in tropical rainforests (forests with an altitude lower than 1000 meters).

The abundance and richness of phyllostomids could change according to altitude, since higher species richness and abundance has been reported in lowland forests (Sampaio et al. 2003; Rex et al. 2008), whereas abundance has been observed to decrease in habitats with altitudes above 1000 meters (McCain 2007; Martins et al. 2015; de Carvalho et al. 2019). Therefore, habitat disturbance could have a major impact on phyllostomid populations in highlands compared to lowlands. In addition, the diversity of plants is high in lowlands but

decreases after 1000 meters (Gentry 1988), which explains the sensitivity of phytophagous
bats to habitat disturbance in highland forests. Other studies have shown the importance of
protecting high altitude tropical forests because human activities put ecosystem services at
risk, for example, the protection and purification of freshwater (Martínez et al. 2009;
Armenteras et al. 2011). Our results support this idea and highlight the importance of the
conservation of highland tropical forests.

359

360 5. Conclusions

361 Although some bats are tolerant to habitat disturbance, because these habitats can provide 362 food and refuges, our results do not suggest that they replace the resources provided by a 363 conserved forest. Bats move through the matrix using both anthropic environments and 364 conserved forests overnight (Ripperger et al. 2015). Therefore, conserved forests will always 365 be essential to maintain phyllostomid bat populations of different trophic guilds. We conclude 366 that the response of phyllostomid guilds to human habitat disturbance is complex and does 367 not depend on phylogenetic signal. Sensitivity occurs regardless of disturbance type or forest 368 type. It is advisable to enrich the anthropogenic matrix with forest cover and 369 chiropterochorous or chiropterophilous species to promote colonization by bats and other 370 animals in order to facilitate their functions and ecosystem services (Kunz et al. 2011; Castro-Luna & Galindo-González 2012). 371

372 The literature used in this meta-analysis was limited because some published studies 373 do not describe the type of habitat disturbance. Also, some studies do not use a control site

- 374 (conserved forest) and they do not show a map or coordinates of the sampled sites. We
- 375 highlight the absence of studies in Central America and the Antilles, which is partly due to the
- 376 limitations already mentioned. For future research, it is necessary to have more information
- about phyllostomid species variables, such as body mass, flight strategies, foraging range,
- 378 and type of refuge used. These can be decisive in understanding the complex response of
- 379 phyllostomid guilds to habitat disturbance.
- 380
- 381 Appendix A. Supplementary data
- 382 The supplementary data to this article are available online.

383 **Conflicts of interest:**

384 The authors of this article have no conflict of interest to declare.

385 Funding

- 386 This research did not receive any specific grant from funding agencies in the public,
- 387 commercial, or not-for-profit sectors.
- 388

389 References

- Armenteras D, Rodríguez N, Retana J, Morales M. 2011. Understanding deforestation in montane and
 lowland forests of the Colombian Andes. Regional Environmental Change 11:693–705. Springer Verlag.
- Arnone IS, Trajano E, Pulchério-Leite A, Passos F de C. 2016. Long-distance movement by a great
 fruit-eating bat, Artibeus lituratus (Olfers, 1818), in southeastern Brazil (Chiroptera,
- 395 Phyllostomidae): evidence for migration in Neotropical bats? Biota Neotropica **16**. Programa
- 396 BIOTA/FAPESP. Available from http://www.scielo.br/scielo.php?script=sci_arttext&pid=S1676-

397 06032016000100102&Ing=en&ting=en (accessed July 17, 2019).

- Ávila-Gómez ES, Moreno CE, García-Morales R, Zuria I, Sánchez-Rojas G, Briones-Salas M. 2015.
 Deforestation Thresholds for Phyllostomid Bat Populations in Tropical Landscapes in the Huasteca Region, Mexico. Tropical Conservation Science 8:646–661.
- 401 Baker RJ, Hoofer SR. 2003. Diversification among New World leaf-nosed bats : an evolutionary
 402 hypothesis and classification inferred from digenomic congruence of DNA sequence. Occasional
 403 Papers, Museum of Texas Tech University 203:1–32.
- Bernard E, Fenton BM. 2003. Bat Mobility and Roosts in a Fragmented Landscape in Central
 Amazonia, Brazil. Biotropica **35**:262.
- Blomberg SP, Garland T, Ives AR. 2003. Testing for phylogenetic signal in comparative data:
 behavioral traits are more labile. Evolution 57:717–745.
- Bobrowiec PE, Lemes MR, Gribel R. 2015. Prey preference of the common vampire bat (Desmodus rotundus, Chiroptera) using molecular analysis. Journal of Mammalogy **96**:54–63.
- Castro-Luna A, Galindo-González J. 2012. Enriching agroecosystems with fruit-producing tree species
 favors the abundance and richness of frugivorous and nectarivorous bats in Veracruz, Mexico.
 Mammalian Biology **77**:32–40.
- Castro-Luna A, Sosa V, Castillo-Campos G. 2007. Bat diversity and abundance associated with the
 degree of secondary succession in a tropical forest mosaic in south-eastern Mexico. Animal
 Conservation 10:219–228.
- 416 Cirranello A, Simmons NB, Solari S, Baker RJ. 2016. Morphological Diagnoses of Higher-Level
 417 Phyllostomid Taxa (Chiroptera: Phyllostomidae). Acta Chiropterologica 18:39–71. Museum and
 418 Institute of Zoology, Polish Academy of Sciences.
- Cisneros LM, Fagan ME, Willig MR. 2015. Effects of human-modified landscapes on taxonomic,
 functional and phylogenetic dimensions of bat biodiversity. Diversity and Distributions 21:523–
 533.
- 422 Cleary KA, Waits LP, Finegan B. 2016. Agricultural intensification alters bat assemblage composition 423 and abundance in a dynamic Neotropical landscape. Biotropica **48**:667–676.
- 424 Cortés-Delgado N, Sosa VJ. 2014. Do Bats Roost and Forage in Shade Coffee Plantations? A
 425 Perspective from the Frugivorous BatSturnira hondurensis. Available from
 426 http://dx.doi.org/10.1111/btp.12142.
- Dalsgaard B et al. 2018. Trait evolution, resource specialization and vulnerability to plant extinctions
 among Antillean hummingbirds. Proceedings. Biological sciences / The Royal Society 285.
 Available from http://dx.doi.org/10.1098/rspb.2017.2754.
- 430 Datzmann T, von Helversen O, Mayer F. 2010. Evolution of nectarivory in phyllostomid bats
 431 (Phyllostomidae Gray, 1825, Chiroptera: Mammalia). BMC evolutionary biology **10**:165.
- de Carvalho WD, Martins MA, Esbérard CEL, Palmeirim JM. 2019. Traits that allow bats of tropical
 lowland origin to conquer mountains: Bat assemblages along elevational gradients in the South
 American Atlantic Forest. Journal of biogeography 46:316–331.
- de la Peña-Cuéllar E, Stoner KE, Avila-Cabadilla LD, Martínez-Ramos M, Estrada A. 2012.
 Phyllostomid bat assemblages in different successional stages of tropical rain forest in Chiapas, Mexico. Biodiversity and Conservation 21:1381–1397.
- 438 Delpietro HA, Marchevsky N, Simonetti E. 1992. Relative population densities and predation of the
 439 common vampire bat (Desmodus rotundus) in natural and cattle-raising areas in north-east
 440 Argentina. Preventive Veterinary Medicine **14**:13–20.
- 441 Díaz S, Purvis A, Cornelissen JHC, Mace GM, Donoghue MJ, Ewers RM, Jordano P, Pearse WD.
 442 2013. Functional traits, the phylogeny of function, and ecosystem service vulnerability. Ecology
 443 and evolution 3:2958–2975.
- 444 Dirzo R, Young HS, Galetti M, Ceballos G, Isaac NJB, Collen B. 2014. Defaunation in the

445 Anthropocene. Science **345**:401–406.

- 446 Dumont ER, Samadevam K, Grosse I, Warsi OM, Baird B, Davalos LM. 2014. Selection for
 447 mechanical advantage underlies multiple cranial optima in new world leaf-nosed bats. Evolution;
 448 international journal of organic evolution 68:1436–1449.
- 449 Durán AA, Pérez SC. 2015. Ensamblaje de murciélagos (Mammalia: Chiroptera) en dos zonas del 450 departamento de Sucre, Colombia. Bat assemblages (Mammalia: Chiroptera). Acta Zoológica 451 Mexicana (n.s) **31**. Available from http://dx.doi.org/10.21829/azm.2015.3131038.
- 452 Edgar RC. 2004. MUSCLE: a multiple sequence alignment method with reduced time and space 453 complexity. BMC bioinformatics **5**:113.
- 454 Egger M, Smith GD, Schneider M, Minder C. 1997. Bias in meta-analysis detected by a simple, 455 graphical test. BMJ **315**:629–634.
- 456 Esbérard CEL, Godoy MSM, Renovato L, Carvalho WD. 2017. Novel long-distance movements by
 457 Neotropical bats (Mammalia: Chiroptera: Phyllostomidae) evidenced by recaptures in
 458 southeastern Brazil. Studies on neotropical fauna and environment 52:75–80. Taylor & Francis.
- 459 FAO. 2012. Global ecological zones for FAO forest reporting: 2010 Update. FAO: Rome, Italy.
- Farneda FZ, Meyer CFJ, Grelle CEV. 2019. Effects of land □ use change on functional and taxonomic
 diversity of Neotropical bats. Biotropica 28:784.
- Farneda FZ, Rocha R, López-Baucells A, Sampaio EM, Palmeirim JM, Bobrowiec PED, Grelle CEV,
 Meyer CFJ. 2018. Functional recovery of Amazonian bat assemblages following secondary forest
 succession. Biological Conservation 218:192–199.
- Fenton MB, Acharya L, Audet D, Hickey MBC, Merriman C, Obrist MK, Syme DM, Adkins B. 1992.
 Phyllostomid Bats (Chiroptera: Phyllostomidae) as Indicators of Habitat Disruption in the Neotropics. Biotropica 24:440.
- Ferreira CMM, Fischer E, Pulchério-Leite A. 2010. Bat fauna in urban remnants of Cerrado in Campo
 Grande, Mato Grosso do Sul. Biota Neotropica **10**:155–160. SciELO Brasil.
- Freeman PW. 2000. Macroevolution in Microchiroptera: recoupling morphology and ecology with
 phylogeny. Mammalogy Papers: University of Nebraska State Museum:8.
- Galindo-González J, Sosa VJ. 2003. Frugivorous bats in isolated trees and riparian vegetation
 associated with human-made pastures in a fragmented tropical landscape. The Southwestern
 naturalist 48:579–590. BioOne.
- García-Morales R, Badano EI, Moreno CE. 2013. Response of neotropical bat assemblages to human
 land use. Conservation biology: the journal of the Society for Conservation Biology 27:1096–
 1106.
- 478 Gentry AH. 1988. Changes in plant community diversity and floristic composition on environmental
 479 and geographical gradients. Annals of the Missouri Botanical Garden. Missouri Botanical
 480 Garden:1–34. JSTOR.
- 481 Gonçalves F, Fischer E, Dirzo R. 2017. Forest conversion to cattle ranching differentially affects
 482 taxonomic and functional groups of Neotropical bats. Biological Conservation 210:343–348.
- Harmon LJ, Weir JT, Brock CD, Glor RE, Challenger W. 2008. GEIGER: investigating evolutionary
 radiations. Bioinformatics 24:129–131.
- 485 Harrison F. 2011. Getting started with meta-analysis. Methods in Ecology and Evolution 2:1–10.
- Heled J, Drummond AJ. 2010. Bayesian inference of species trees from multilocus data. Molecular
 biology and evolution 27:570–580.
- Jones G, Jacobs DS, Kunz TH, Willig MR, Racey PA. 2009. Carpe noctem: the importance of bats as
 bioindicators. Endangered Species Research 8:93–115.
- Jones P, Hämsch F, Page RA, Kalko EKV, Teague O'Mara M. 2017. Foraging and Roosting
 Behaviour of the Fringe-Lipped Bat, Trachops cirrhosus, on Barro Colorado Island, Panamá. Acta
- 492 Chiropterologica **19**:337–346.

- Kalka MB, Smith AR, Kalko EKV. 2008. Bats Limit Arthropods and Herbivory in a Tropical Forest.
 Science 320:71–71.
- Kalko E, Friemel D, Handley CO, Schnitzler H-U. 1999. Roosting and Foraging Behavior of Two
 Neotropical Gleaning Bats, Tonatia silvicola and Trachops cirrhosus (Phyllostomidae). Biotropica
 31:344–353.
- Kalko E, Ueberschaer K, Dechmann D. 2006. Roost Structure, Modification, and Availability in the
 White-throated Round-eared Bat, Lophostoma silvicolum (Phyllostomidae) Living in Active
 Termite Nests. Biotropica 38:398–404.
- 501 Kembel SW, Cowan PD, Helmus MR, Cornwell WK, Morlon H, Ackerly DD, Blomberg SP, Webb CO. 502 2010. Picante: R tools for integrating phylogenies and ecology. Bioinformatics **26**:1463–1464.
- 503 Kingman JFC. 1982. The coalescent. Stochastic Processes and their Applications **13**:235–248.
- 504 Klingbeil BT, Willig MR. 2009. Guild-specific responses of bats to landscape composition and 505 configuration in fragmented Amazonian rainforest. Journal of Applied Ecology **46**:203–213.
- 506 Kunz TH, de Torrez EB, Bauer D, Lobova T, Fleming TH. 2011. Ecosystem services provided by bats. 507 Annals of the New York Academy of Sciences **1223**:1–38.
- Martínez ML, Pérez-Maqueo O, Vázquez G, Castillo-Campos G, García-Franco J, Mehltreter K,
 Equihua M, Landgrave R. 2009. Effects of land use change on biodiversity and ecosystem
 services in tropical montane cloud forests of Mexico. Forest ecology and management 258:1856–
 1863.
- Martins MA, De Carvalho WD, Dias D, De S. França D, De Oliveira MB, Peracchi AL. 2015. Bat
 Species Richness (Mammalia, Chiroptera) Along an Elevational Gradient in the Atlantic Forest of
 Southeastern Brazil. Acta Chiropterologica 17:401–409.
- McCain CM. 2007. Could temperature and water availability drive elevational species richness
 patterns? A global case study for bats. Global ecology and biogeography: a journal of
 macroecology 16:1–13.
- 518 Medellín RA, Equihua M, Amin MA. 2000. Bat Diversity and Abundance as Indicators of Disturbance 519 in Neotropical Rainforests. Conservation Biology **14**:1666–1675.
- Medellin RA, Rivero M, Ibarra A, de la Torre JA, Gonzalez-Terrazas TP, Torres-Knoop L, Tschapka M.
 2018. Follow me: foraging distances of Leptonycteris yerbabuenae (Chiroptera: Phyllostomidae)
 in Sonora determined by fluorescent powder. Journal of mammalogy 99:306–311. Narnia.
- 523 Münkemüller T, Lavergne S, Bzeznik B, Dray S, Jombart T, Schiffers K, Thuiller W. 2012. How to 524 measure and test phylogenetic signal. Methods in Ecology and Evolution **3**:743–756.
- Murillo-García OE, Bedoya-Durán MJ. 2014. Distribución y abundancia de murciélagos en bosques
 con diferente grado de intervención en el Parque Nacional Natural Gorgona (Colombia). Revista
 de Biología Tropical 62:419.
- 528 Muscarella R, Fleming TH. 2007. The role of frugivorous bats in tropical forest succession. Biological 529 reviews of the Cambridge Philosophical Society **82**:573–590.
- Nakagawa S, Noble DWA, Senior AM, Lagisz M. 2017. Meta-evaluation of meta-analysis: ten
 appraisal questions for biologists. BMC biology 15:18.
- 532 Newbold T et al. 2015. Global effects of land use on local terrestrial biodiversity. Nature **520**:45–50.
- Newbold T, Bentley LF, Hill SLL, Edgar MJ, Horton M, Su G, Şekercioğlu ÇH, Collen B, Purvis A.
 2020. Global effects of land use on biodiversity differ among functional groups. Functional ecology. Available from https://onlinelibrary.wiley.com/doi/abs/10.1111/1365-2435.13500.
- Ochoa J. 2000. Efectos de la Extraccion de Maderas sobre la Diversidad de Mamiferos Pequenos en
 Bosques de Tierras Bajas de la Guayana Venezolana. Biotropica 32:146–164.
- Pardo LE, Campbell MJ, Cove MV, Edwards W, Clements GR, Laurance WF. 2019. Land
 management strategies can increase oil palm plantation use by some terrestrial mammals in
 Colombia. Scientific reports 9:7812.

- Peters SL, Malcolm JR, Zimmerman BL. 2006. Effects of selective logging on bat communities in the
 southeastern Amazon. Conservation biology: the journal of the Society for Conservation Biology
 20:1410–1421.
- 544 Potapov P et al. 2017. The last frontiers of wilderness: Tracking loss of intact forest landscapes from 545 2000 to 2013. Science advances **3**:e1600821.
- 546 Prone B, Zanon CMV, Benedito E. 2012. Bats (Chiroptera, Phyllostomidae) in the urbanized area in
 547 South of Brazil. Acta Scientiarum. Biological Sciences 34:155–162. Universidade Estadual de
 548 Maringa.
- Quinto-Mosquera J, Moreno-Amud LA, Mosquera-Mosquera S, Mantilla-Meluk H, Jimenez-Ortega
 AM. 2013. Cambios en el ensamblaje de murciélagos filostómidos producto de la perturbación en el distrito Alto Atrato-San Juan, Chocó, Colombia. Revista Institucional Universidad Tecnológica del Chocó. Investigación, Biodiversidad y Desarrollo **32**:121–132.
- 553R Core Team (2018). R: A language and environment for statistical computing. R Foundation for554Statistical Computing, Vienna, Austria. Available from https://www.R-project.org/.
- Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA. 2018. Posterior Summarization in Bayesian
 Phylogenetics Using Tracer 1.7. Systematic Biology 67:901–904.
- Revell LJ. 2012. phytools: an R package for phylogenetic comparative biology (and other things).
 Methods in Ecology and Evolution 3:217–223.
- Rex K, Czaczkes BI, Michener R, Kunz TH, Voigt CC. 2010. Specialization and omnivory in diverse
 mammalian assemblages. Écoscience 17:37–46.
- Rex K, Kelm DH, Wiesner K, Kunz TH, Voigt CC. 2008. Species richness and structure of three
 Neotropical bat assemblages. Biological journal of the Linnean Society. Linnean Society of
 London 94:617–629. Narnia.
- Ripperger SP, Kalko EKV, Rodríguez-Herrera B, Mayer F, Tschapka M. 2015. Frugivorous bats
 maintain functional habitat connectivity in agricultural landscapes but rely strongly on natural
 forest fragments. PloS one **10**:e0120535.
- Rojas D, Vale A, Ferrero V, Navarro L. 2011. When did plants become important to leaf-nosed bats?
 Diversification of feeding habits in the family Phyllostomidae. Molecular ecology 20:2217–2228.
- Saldaña-Vázquez RA, Schondube JE. 2016. La masa corporal explica la dominancia de Artibeus
 (Phyllostomidae) en ambientes urbanos. Pages 23–33 Memorias en Extenso del I Congreso de
 Fauna Nativa en Medios Antropizados (A. Ramírez-Bautista, y R. Pineda-López, eds.).
 CONACYT-UAQ, México.
- Saldaña-Vázquez RA, Sosa VJ, Hernández-Montero JR, López-Barrera F. 2010. Abundance
 responses of frugivorous bats (Stenodermatinae) to coffee cultivation and selective logging
 practices in mountainous central Veracruz, Mexico. Biodiversity and Conservation 19:2111–2124.
- Sampaio EM, Kalko EKV, Bernard E, Rodríguez-Herrera B, Handley CO. 2003. A Biodiversity
 Assessment of Bats (Chiroptera) in a Tropical Lowland Rainforest of Central Amazonia, Including
 Methodological and Conservation Considerations. Studies on neotropical fauna and environment
 38:17–31. Taylor & Francis.
- Schneider CA, Rasband WS, Eliceiri KW. 2012. NIH Image to ImageJ: 25 years of image analysis.
 Available from https://www.ncbi.nlm.nih.gov/pubmed/22930834.
- 582 Shi JJ, Rabosky DL. 2015. Speciation dynamics during the global radiation of extant bats. Evolution; 583 international journal of organic evolution **69**:1528–1545.
- Socolar JB, Sandoval EHV, Wilcove DS. 2019. Overlooked biodiversity loss in tropical smallholder
 agriculture. Conservation biology: the journal of the Society for Conservation Biology. Available
 from http://dx.doi.org/10.1111/cobi.13344.
- 587 Suchard MA, Lemey P, Baele G, Ayres DL, Drummond AJ, Rambaut A. 2018. Bayesian phylogenetic 588 and phylodynamic data integration using BEAST 1.10. Virus evolution **4**:vey016.

- 589 Venter O et al. 2016. Sixteen years of change in the global terrestrial human footprint and implications 590 for biodiversity conservation. Nature communications **7**:12558.
- 591 Viechtbauer W. 2010. Conducting Meta-Analyses in R with the metafor Package. Journal of Statistical 592 Software **36**. Available from http://dx.doi.org/10.18637/jss.v036.i03.
- 593 Villalobos F, Arita HT. 2010. The diversity field of New World leaf-nosed bats (Phyllostomidae). Global 594 Ecology and Biogeography **19**:200–211.
- Voss RS, Fleck DW, Strauss RE, Velazco PM, Simmons NB. 2016. Roosting Ecology of Amazonian
 Bats: Evidence for Guild Structure in Hyperdiverse Mammalian Communities. American Museum
 Novitates 3870:1–43.
- Williams-Guillén K, Perfecto I. 2010. Effects of Agricultural Intensification on the Assemblage of Leaf Nosed Bats (Phyllostomidae) in a Coffee Landscape in Chiapas, Mexico. Biotropica 42:605–613.
- 600 Willig MR, Presley SJ, Plante J-L, Bloch CP, Solari S, Pacheco V, Weaver SC. 2019. Guild-level
- responses of bats to habitat conversion in a lowland Amazonian rainforest: species composition
 and biodiversity. Journal of Mammalogy **100**:223–238.

603 Figures



Figure 1. Map of the locations of the studies used for the generation of the database. Each

- point indicates the number of cases that contributed to the analysis. Boxes on the right show
- a close up of the areas marked as a and b on the main map.



Figure 2. Phylogenetic reconstruction of the Phyllostomidae family using COI, RAG2, and CTBY genes. The white-black gradient in each branch is related to the proportion of the abundance of each observed species. The figure shows that there is no phylogenetic signal in the response of bats to habitat disturbance.

611 Tables

- Table 1. Definition of the seven types of habitat disturbance and the number of cases for each
- 613 one.

Disturbance type	Definition	Number of cases (<i>k</i>)
Crops	All land used for growing a product regardless of its type and intensity. These were subclassified into: monoculture, polyculture, agroforestry, and urban crops.	262
Grassland	Land covered to a greater extent by grass in an induced manner and for livestock purposes.	21
Grassland-Crops	Mixed use of land by induced grassland for livestock and crops.	20
Logging	Forest intervened by selective logging.	107
Secondary forest	Secondary vegetation where, after the removal of the forest, there has been a process of natural succession. The number of years the process takes without any type of human intervention was taken.	292
Tourism	Forest intervened with the presence of trails and buildings for tourism purposes.	7
Urban zone	Sites with vegetation within the urban matrix, such as home gardens and urban parks.	54

- Table 2. Response of frugivorous (F), nectarivorous (N), and insectivorous-frugivorous-615
- nectarivorous (INF) bats to different types of habitat disturbance. 616

Trophic guild (<i>k</i>)	QM ^a (d.f.)	Disturbance type	Estimate ^b	Cl ^c	Р
F (462)	1019.5690 (7)	Logging ^d	0.2029	0.0932-0.312	0.0003
		Tourism ^e	0.751	0.634-0.869	<0.0001
		Secondary forest ^e	0.556	0.459-0.653	<0.0001
		Crops ^e	0.558	0.461-0.656	<0.0001
		Grassland ^d	0.215	0.115-0.315	<0.0001
		Grassland-crops	0.221	0-0.662	0.325
		Urban zone ^d	0.3202	0.219-0.421	<0.0001
		Global analysis	0.4808	0.454-0.507	<0.0001
N (41)	413.7397 (6)	Logging	0.552	0.418-0.687	<0.0001
		Tourism	0.823	0.451-1	<0.0001
		Secondary forest ^e	0.785	0.686-0.885	<0.0001
		Crops ^e	0.641	0.522-0.7605	<0.0001
		Grassland-crops	0.5	0.0799-0.9201	0.0197
		Urban zone ^d	0.219	0-0.452	0.0651
		Global analysis	0.566	0.478-0.653	<0.0001
INF (26)	504.4064 (6)	Logging ^a	0.226	0.0683-0.383	0.005
		Secondary forest ^e	0.813	0.712-0.913	<0.0001
		Crops ^d	0.359	0.245-0.473	<0.0001
		Grassland ^d	0.129	0-0.4707	0.4577
		Grassland-crops ^e	0.962	0.839-1	<0.0001
		Urban zone	0.363	0.0013-0.725	0.0492
		Global analysis	0.473	0.349-0.596	<0.0001

^aCochran's Q index value for the heterogeneity test 617

^bis the proportion of bat abundance estimated for each model. ^cConfidence interval of the estimate value 618

- 619
- ^dare sensitive and do not transpose their confidence intervals with ^e 620

621 ^eare tolerant and do not transpose their confidence intervals with ^d

622 Table 3. Response of frugivorous (F) and frugivorous-nectarivorous (FN) bats to disturbance

623 type in different forest types: Tropical Rainforest (TRF), Tropical Dry Forest (TDF), Tropical

624 Moist Forest (TMF), Tropical Mountain System (TMS), and Subtropical Mountain System

625 (STMS).

Trophic			Forest			
guild (<i>k</i>)	QM ^a (d.f.)	Disturbance type	type	Estimate	Cl ^c	р
E (462)	1010 5000 (15)	Sacandary foract	TDE	0 576	0 241 0 911	4 0001
F (402)	1213.5333 (15)	Secondary lorest		0.576	0.341-0.811	<.0001
				0.284	0-0.617	0.0943
			TMS ^u	0.233	0-0.469	0.0527
			TRF ^e	0.627	0.515-0.738	<.0001
		Crops	STMS	0.295	0-0.627	0.0818
			TMS	0.587	0.349-0.826	<.0001
			TRF	0.588	0.476-0.699	<.0001
		Grassland	TMS	0.0264	0-0.263	0.827
			TRF	0.144	0.028-0.2603	0.015
		Urban zone	TMF ^d	0.0451	0-0.377	0.7901
			TMS ^d	0.068	0-0.4015	0.6894
			TRF ^e	0.6061	0.4098-0.8024	<.0001
		Global analysis		0.4808	0.454-0.5074	<.0001
FN (52)	223.4907 (12)	Secondary forest	TDF ^e	0.7304	0.485-0.975	<.0001
		-	TMF	0.75	0.0594-1	0.0333
			TMS ^d	0.0385	0-0.388	0.829
			TRF ^e	0.697	0.563-0.831	<.0001
		Crop	STMS	0.823	0.454-1	<.0001
			TMS ^d	0.116	0-0.446	0.489
			TRF ^e	0.628	0.493-0.762	<.0001
		Grassland	TMS	0.0385	0-0.395	0.832
			TRF	0.168	0-0.3809	0.121
		Global analysis		0.524	0.443-0.6043	<.0001

626 ^aCochran's Q index value for the heterogeneity test

^bis the proportion of bat abundance estimated for each model.

- 628 ^cConfidence interval of the estimate value
- 629
- ^dare sensitive and do not transpose their confidence intervals with ^e ^eare tolerant and do not transpose their confidence intervals with ^d 630